Life Science Journal

Websites: http://www.lifesciencesite.com http://www.sciencepub.net

Emails: editor@sciencepub.net sciencepub@gmail.com



Chemical composition and *in situ* digestion kinetics of urea-molasses treated fallen leaves of *Quercus rugosa*

Carrillo-Muro Octavio¹, Ramírez Roque Gonzalo¹, Hernández-Briano Pedro², López-Carlos Marco Antonio², González-Ronquillo Manuel³, Aguilera-Soto Jairo Iván^{2,*}

¹Facultad de Ciencias Biológicas, Universidad Autónoma de Nuevo León, San Nicolás de los Garza, Nuevo León, 66450, México

²Unidad Académica de Medicina Veterinaria y Zootecnia, Universidad Autónoma de Zacatecas, Gral. Enrique Estrada, Zacatecas, 98560, México

³Facultad de Medicina Veterinaria y Zootecnia, Universidad Autónoma del Estado de México, Toluca, 50090,

México.

aguileraivan@yahoo.com.mx

Abstract: The aim of the study was to determine the effect of the use of urea (0, 2, 4 and 6%) and or molasses (0, 15, 30 and 45%) on the nutritive value of leaves of *Quercus rugosa* Née The *in situ* digestibility of dry matter (ISDDM) was also estimated. The metabolizable energy (ME), net energy of lactancy (NEI), the organic matter digestion coefficient (OMD) and short chain volatile fatty acids (SCFA) were determined from data of the *in vitro* gas production. Leaves were collected from three sites: site 1 was located in Tequila, county of the state of Jalisco; site 2 was situated in Teul de González Ortega county of the state of Zacatecas, México, and site 3 in San Cristobal de La Barranca county of the state of Jalisco, México. In general, the organic matter, neutral detergent fiber and acid detergent fiber contents diminished as the levels of molasses increased in diets. Conversely, crude protein, nonfiber carbohydrates, ash, ME, NEI and SCFA contents and ISDMD augmented as molasses increased. In general, when urea and molasses were higher, the fractions **a**, **b**, **a**+**b** of the *in situ* digestibility were higher. In all the molasses mixtures, with addition of urea at 4 or 6% had higher chemical composition and *in situ* digestibility.

[Carrillo-Muro O, Ramírez RG, Hernández-Briano P, López-Carlos MA, González-Ronquillo M, Aguilera-Soto JJ **Chemical composition and** *in situ* digestion kinetics of urea-molasses treated fallen leaves of *Quercus rugosa*. *Life Sci J* 2022;19(5):20-27]. ISSN 1097-8135 (print); ISSN 2372-613X (online). <u>http://www.lifesciencesite.com</u>. 4. doi:10.7537/marslsj190522.04.

Keywords: Quercus rugosa; chemical composition; in situ digestibility; urea; molasses.

Websites: http://www.lifesciencesite.com http://www.sciencepub.net

Emails: editor@sciencepub.net sciencepub@gmail.com

1. Introduction

Recently, the use of forest residues as foods for ruminants has been focused mainly in find alternatives for the utilization of feed sources of low cost. The great amounts of fallen leaves may be considered as biomass that is ignored as animal feed, and may be considered as an alternative source of forage for ruminants (De la Paz et al., 1998). In this region, the fallen leaves of *Q. rugosa* may be used for food source for ruminants at the end of winter when defoliation occurs (Valencia, 2004). However, fallen leaves from Quercus have high concentrations of structural carbohydrates and low crude protein content and low of dry matter digestibility. Thus, to improve the rumen digestion of straw fed animals in terms of the availability of nonfiber carbohydrates and ammonia, supplementation of urea and molasses in the form of block or as liquid feed is often suggested (Reis et al., 1995; Sahoo et al., 2004). Other studies have reported beneficial effects of urea and molasses treatment on rumen degradability of wheat, oat and barley straws (Van Thu and Uden, 2000; Souza y Dos Santos, 2002; Montiel et al., 2012).

In situ digestion parameters: the soluble fraction **a**, the degradable fraction **b**, the potential degradable fraction $\mathbf{a}+\mathbf{b}$ and rate constant \mathbf{c} are important characteristics of forage digestion in ruminants. Such parameters can be used to predict the nutritive value more accurately and compare the utility of forages in the diets for ruminants (Ørskov, 2000). Therefore, this study was carried out with the aim to determine the effect of the inclusion of urea (0, 2, 4 and 6%) and molasses (0, 15, 30 and 45%) to the fallen leaves of Q. rugosa Née on the chemical composition and in situ digestion parameters. It was hypothesized that addition of the mixture of a nitrogen source (urea) and a soluble carbohydrate (molasses) on fallen leaves of O. rugosa will improve its solubility of structural carbohydrates and crude protein.

2. Material and Methods

2.1. Study sites

The collection sites were characterized by a similar edaphic pattern, ground elevations, climate conditions and vegetation (CONAFOR, 2009). Predominant soil type was extrusive igneous rock, and phaeozem leptosol. The climate of the region is sub humid warm with summer rains. The annual mean rainfall is about 600 mm. The collection site 1 was located at 21°15′07.30″ latitude N and 103°39′48.12″ longitude W, in the municipality of



Tequila, Jalisco, Mexico; at an altitude of 1,617 m. The collection site 2 was located at 21°17'46.02" latitude N and 103° 34'52.03" longitude W, in the municipality of Teul de Gonzalez Ortega, Zacatecas, Mexico; at an altitude of 1,997 m. The collection site 3 was located at 21°11'30.14" latitude N and 103°35'59" longitude W, in the municipality of San Cristobal de la Barranca, Jalisco, Mexico; at an altitude of 1,738 m (INEGI, 2009abc).

2.2. Harvest procedure and sample handling

Sampling of fallen leaves of Quercus was carried out in two years (2013 and 2014), during the second week of March. Leaves were collected in a plot of about 50 m \times 50 m randomly located in each collection site. Leaf litter was collected by hand and keep in polyethylene bags of 1.4 m \times 1.0 m. The bags were transported to the laboratory and stored at 22 °C. Grinding of leaf litter was achieved with a mill (Azteca brand No. 24) equipped with a 2" screen and powered by a tractor at 105 hp engine and 90.4 hp PTO (New Holland 7610S 4WD Double).

2.3 Ensiled the urea and molasses with the fallen leaves of Quercus

There were 16 treatments (mixtures) composed by fallen leaves of Ouercus with four levels of urea (0, 2, 4 and 6%) and four levels of molasses of 85 °Brix (0, 15, 30 and 45%). Grinded leaves (1.5 kg) were mixed with the ingredients for each treatment: first diluted urea with 10% of water was added to leaves, then the molasses were included and completely homogenized the three components of the treatment mixtures. Each treatment mixture were ensiled in cylindrical laboratory silos made by polyvinylchloride of 5 cm in diameter and 30 cm long and sealed in each extreme of the silo. There were four experimental silos for each treatment mixture. After a period of 28 days, the silos that were placed at 22 °C, were opened and mixtures were dried at 60 °C for 48 hours in a air circulating oven. Then were grinded in a Wiley mill to pass a 1mm screen. Finally, grinded samples of each treatment mixture were dried again for 24 hours at 60 °C for chemical and digestion analyses.

2.4 Chemical analyses

By triplicate each treatment mixture, was analyses to determine organic matter (OM), ash (AOAC, 1995), neutral detergent fiber (NDF) and acid detergent fiber (ADF; Van Soest *et al.*, 1991). Hemicellulose (NDF - ADF) and OM (DM - ash) were determined by difference. Nonfiber carbohydrate (NFC) were calculated by the equation of Sniffen *et al.* (1992): NFC (%) = DM - (CP + NDF + EE + ash). The Dumas method, (AOAC, 1995) was used for the determination of crude protein (CP) with a FP-528 LECO apparatus. The ether extract (EE) was determined using the extractor of Ankom^{xt15}. The concentration of condensed tannins (CT) was estimated using the butanol-HCl method (Makkar, 2005).

2.5 In vitro gas production

Gas production was determined by the *in vitro* procedure proposed by Theodorou *et al.* (1994). Briefly, 700 \pm 10 mg of DM of each sample were introduced into fermentation units (FU) of 120 ml, mixed with 90 ml of a buffer solution gassed with CO₂ and stored under refrigeration (4°C). After 24 hours, 10 ml of ruminal fluid were also added to the FU. Finally, the FU were kept in a water bath at 39 °C. The gas production was measured using a pressure transducer (SPER SCIENTIFIC, 840065 gauge with a pressure gauge TA87199). The pressure of gas produced was determined in PSI (Pounds-force per Square Inch) at 24 h of incubation. Samples were assayed by quadruplicate.

The ruminal fluid was obtained from three ruminally cannulated sheep fed a diet (12.8% of CP, 2.34 Mcal/kg of ME and 64.60% of TDN NRC, 2007) containing 82% hay (50% alfalfa and 50% wheat straw), 17% concentrate (63% corn cracked, 25% of cottonseed meal, 5.5% of limestone, 5.5% of monocalcium phosphate), and 1 % of vitamins and microminerals premix. Feed was provided *ad libitum*, twice daily at 0800 and 1600 h, with free access to water.

To correct for contamination with rumen contents three FU without substrate were used as whites. In addition, three FU with bean straw, three with bean straw + corn cracked and three with concentrated were used as standards. Pressure readings (PSI) were converted to volume (ml) using a preset linear regression between recorded in such units and inoculated known volumes of air at the same temperature incubation pressures such as: Volume= (Pressure-0.4009) / 0.3853; R = 0.9959; SEM = 0.0522; and n= 72.

Since the *in vitro* production of gas is proportional to the DM degraded, the net yield of gas at 24 h (ml/700 mg) incubation of the substrate was used to calculate the metabolizable energy (ME) and IVOMD using the equations proposed by Menke and Steingas (1998), as follows:

ME, Mcal/kg⁻¹ DM = $(2.20+0.136 \times GP_{24h}+0.057 \times CP+0.0029 \times EE^2)/4.184$

IVOMD, % = $14.88+0.889 \times GP_{24h}$ + $0.045 \times CP + 0.0651 \times XA$.

The net energy of lactation (NEl) was also calculated by the equation of Abas *et al.* (2005):

 $NEI (Mcal/kg^{-1} DM) = (0.115 \times GP_{24h} + 0.0054 \times CP + 0.014 \times EE - 0.0054 \times XA - 0.36)/4.184$

The short chain fatty acids (SCFA) were calculated using the equation of Makkar (2005):

SCFA (mmol) = $0.0222 \times GP_{24h} - 0.00425$

Where GP= net gas production at 24 h (ml/700 mg); CP= crude protein; EE = ether extract; and XA = ash content.

2.5. Dynamics of in situ degradability

The technique of nylon bag (Ørskov, 2000) was used to assess the degradability of MS. Treatment mixtures (5 g) were incubated in the rumen of five rams equipped with ruminal cannula, for which they were suspended in the ventral part of the rumen for 0, 6, 12, 24, 48, 72 and 96 h of incubation. The samples were introduced in the reverse order of incubation time to be later removed all together. The zero time bags were introduced and removed immediately in order to wet with ruminal fluid (Nocek and Russell, 1988). To determine the parameters of *in situ* degradability and passage rate, the data obtained were processed with the Neway (Cheng, 1997) computer program, applying the equation proposed by Ørskov and McDonald (1979): $p = a + b (1 - e^{-ct}),$

Where, $\mathbf{p} = \%$ degradation of dry matter (DM) at time t; $\mathbf{a} =$ soluble or rapidly degradable fraction; $\mathbf{b} =$ fraction insoluble or slowly degradable in time t; $\mathbf{e} =$ natural logarithm; $\mathbf{c} =$ degradation constant b; $\mathbf{t} =$ time of incubation. It follows that $\mathbf{a} + \mathbf{b} =$ maximum potential food degradability; and effective degradation, which corresponds to the degradation potential ($\mathbf{a} + \mathbf{b}$) adjusted by the effect of passage rate (\mathbf{k}), by the relationship $\mathbf{p} = \mathbf{a} + (\mathbf{b} \times \mathbf{c})/(\mathbf{c} + \mathbf{K}\mathbf{p})$ was calculated for $\mathbf{K}\mathbf{p}$ values of 3% h⁻¹, corresponding to moderate production diets.

2.6. Statistical analyses

Data was analyzed as a one way analysis of variance in a 4 x 4 factorial arrangement considering the urea (0, 2, 4 and 6%) and molasses (0, 15, 30 and 45%) levels as the main effects. The GLM procedure of the SAS statistical software (SAS/STAT® User's Guide (8.1Edition), SAS Inst. Inc., Cary, NC, USA (SAS, 2000) was used to compute the data. When significant effects were observed, a comparison of means with the Tukey method using the MEANS statement was made. P values observed were considered different if P<0.05.

3. Results and discussion

3.1. Chemical composition

The OM content was higher (P<0.01) in treatments only with urea (Table 1). Araiza-Rosales *et al.* (2013) reported similar response in corn silage with molasses at 0, 5 y 10%. The CP content significantly increased as the addition of urea and molasses increased in fallen leaves of Quercus being the higher treatment with 45% molasses and 6% urea (Table 1). Conversely, Vallejo-Solís (1995) argued that The CP content in fallen leaves of native trees and shrubs was reduced as molasses augmented in diets. In general, the EE content significantly the leaf litter was reduced as urea and molasses was increased.

The NDF and ADF of fallen leaves were significantly reduced as the addition of urea and molasses increased. Araiza-Rosales et al. (2013) reported similar tendency in reduction of structural carbohydrates in corn silage treated with molasses at 0, 5 y 10%. With exception of the treatment with 0%molasses, all others (15, 30 and 45%) augmented as urea increased. The NFC content significantly reduced as the addition of urea and molasses increased. In general, as the percentage of urea and molasses increased the ME, NEl and SCFA contents in treated fallen leaves, significantly augmented (Table 1). Similar findings were reported by Shultz et al. (1971) who reported increments of SCFA in rumen samples of beef cattle fed with Panicum maximum hay treated with 2.4% of molasses.

3.2. Digestion parameters

The soluble fraction **a** of fallen leaves significantly augmented as the addition of urea and molasses increased in fallen leaves of Quercus (Table 2). Similar findings were observed by Araiza-Rosales et al. (2013) with corn silage added with 0, 5 and 10% molasses. They also reported that with 10% molasses the fraction **a** was significantly higher than other levels of molasses. In this study, the fraction **b** significantly varied among treatments being higher when fallen leaves were added with 6% urea and 45% of molasses. The rate constant of degradation c of the dry matter of the fallen leaves of Quercus significantly augmented as the addition of urea and molasses augmented (Table 2). The potential degradability **a**+**b** of the dry matter of fallen leaves treated with urea and molasses varied significantly. This fraction in general augmented as the levels of urea and molasses increased. Conversely, Vallejo-Solís (1995) reported that the potential degradability of silages based on tree and shrub fodders was reduced as molasses as the levels of molasses were increased. In this study, the fraction **c** was significantly different among treatments. Higher digestion rates were achieved in leaf litter samples with molasses and 4 and 6% of urea (Table 2). In concordance with this study, Souza and Dos Santos (2002) reported that the rate of digestion of barley straw increased as the level of urea increased from 0 to 6%. Conversely, Pinto-Hernández *et al.* (1994) mentioned that the digestion parameters do not varied in the forage of *Panicum maximum* treated with or without molasses and urea.

Life Science Journal

Websites: http://www.lifesciencesite.com http://www.sciencepub.net

Emails: editor@sciencepub.net sciencepub@gmail.com



Content		0% mc	lasses		15% molasses				30% molasses				45% molasses				
	Urea, %																
	0	2	4	6	0	2	4	6	0	2	4	6	0	2	4	6	SEM
ОМ, %	92.2 ^d	93.7 ^b	93.9 ^{ab}	94.2ª	92.1 ^{de}	92.7°	92.7°	92.8°	90.7 ⁱ	91.3 ^g	91.7 ^f	91.9 ^{ef}	91.0 ^{hi}	91.1 ^{hg}	90.9 ^{hi}	91.1 ^{hg}	0.05
Ash, %	7.8°	6.3 ^g	6.0 ^{gh}	5.8 ^h	8.0 ^{de}	7.33 ^f	7.3 ^f	7.2 ^f	9.3ª	8.7°	8.2 ^d	8.0 ^{de}	9.0 ^{ab}	8.9 ^{bc}	9.1 ^{ab}	8.9 ^{bc}	0.05
Crude protein, %	1.9 ^k	7.5 ^h	11.7°	17.2°	2.7 ^j	7.9 ^h	12.0 ^e	17.8 ^b	3.6 ⁱ	8.5 ^g	13.2 ^d	17.9 ^{ab}	8.9 ^g	9.4 ^f	13.4 ^d	18.2ª	0.08
Ether extract, %	1.6 ^a	1.4 ^{bc}	1.1 ^{def}	0.9 ^f	1.5 ^{ab}	1.3 ^{bcd}	1.0 ^{ef}	$1.1^{\rm f}$	1.4 ^{bc}	1.3 ^{bcd}	1.2 ^{cde}	1.2 ^{cde}	1.3 ^{bc}	1.2 ^{cde}	1.2 ^{cde}	1.2 ^{cde}	0.04
NDF, %	67.7 ^{ab}	70.3ª	69.7ª	69.5ª	63.6 ^{bcd}	62.7 ^{bcde}	67.3 ^{ab}	64.2 ^{bc}	59.1 ^{cde}	58.5 ^{def}	59.8 ^{cde}	58.0 ^{ef}	53.8 ^{fg}	60.1 ^{cde}	57.6 ^{efg}	52.6 ^g	0.9
ADF, %	62.1 ^{ab}	64.1ª	64.7ª	64.8ª	57.9 ^{bc}	56.6 ^{bcd}	60.2 ^{ab}	56.3 ^{bcde}	53.0 ^{cdef}	53.5 ^{cdef}	52.0 ^{cdef}	50.5 ^{ef}	48.6 ^g	51.8 ^{def}	49.8 ^g	44.4 ^g	1.0
Hemicellulose, %	5.6 ^{bcd}	6.2 ^{abcd}	4.9 ^d	4.7 ^d	5.6 ^{bcd}	6.1 ^{abcd}	7.0 ^{abcd}	7.9 ^{ab}	6.1 ^{abcd}	5.0 ^d	7.7 ^{ab}	7.4 ^{abc}	5.2 ^{cd}	8.3ª	7.8 ^{ab}	8.2ª	0.4
NFC, %	20.90 ^{bcd}	14.3 ^{fg}	11.4 ^{fgh}	6.4 ^h	24.1 ^{ab}	20.6 ^{bcd}	12.4 ^{fg}	9.7 ^{hg}	26.7ª	22.8 ^{abc}	17.6 ^{ed}	14.8 ^{ef}	26.8ª	20.3 ^{bcd}	18.9 ^{cde}	19.0 ^{cde}	0.90
ME, MJ kg ⁻¹	3.3 ^h	3.8 ^{gh}	3.8 ^{gh}	4.2 ^{efgh}	4.2 ^{efgh}	4.2 ^{efgh}	4.6 ^{defg}	5.4 ^{cde}	5.0 ^{defg}	50 ^{def}	5.9 ^{bcd}	6.7 ^{abc}	6.9 ^b	6.7 ^{ab}	6.8 ^{ab}	7.5 ^a	0.06
NEl, MJ kg ⁻¹	0.4 ^{fg}	0.4^{fg}	0.4 ^g	0.8 ^{cfg}	1.3 ^{def}	0.8^{efg}	0.8 ^{efg}	2.1 ^{cde}	1.3 ^{def}	1.3 ^{def}	2.1 ^{bcd}	3.3ª	2.9 ^b	2.8 ^{bc}	2.9 ^b	3.8ª	0.05
SCFA, mmol	0.1°	0.1 ^e	0.1 ^e	0.1 ^e	0.3 ^{cde}	02 ^{de}	0.2 ^{de}	0.4 ^{bcd}	0.3 ^{cde}	0.3 ^{cde}	0.4 ^{abc}	0.7 ^b	0.6 ^{ab}	0.6 ^{ab}	0.6 ^{ab}	0.8 ^a	0.05

Table 1. Chemical composition of fallen leaves of Quercus rugosa treated with different levels of urea and molasses

OM = organic matter; NDF = neutral detergent fiber; ADF = acid detergent fiber; NFC = non-fiber carbohydrates; ME = metabolizable energy; NEl = net energy for lactancy; SCFA = short chain fatty acid; SEM = standard error of the mean.

a, b, c, d. f. g h Means in a row with different letter superscripts are different (P<0.05).

Concept		0% mo	olasses		15% molasses				30% molasses				45% molasses				
	Urea, %																
	0	2	4	6	0	2	4	6	0	2	4	6	0	2	4	6	SEM
a, %	18.2 ^g	18.5 ^g	16.5 ^g	23.8 ^f	22.8 ^f	23.2 ^f	27.4 ^e	29.3 ^d	33.6 ^{bcd}	33.6 ^{bcd}	36.0 ^{ab}	33.4 ^{bcd}	33.7 ^{bcd}	35.0 ^{abcd}	36.8 ^a	35.7 ^{abc}	0.50
b, %	13.7 ^{bcd}	13.3 ^{bed}	20.8ª	14.2 ^{bcd}	17.9 ^{ab}	18.2 ^{ab}	18.0 ^{ab}	17.2 ^{ab}	7.9 ^d	8.3 ^d	10.1 ^{cd}	13.5 ^{bcd}	12.6 ^{cd}	13.7 ^{bc}	11.23 ^{cd}	22.3ª	0.90
a+b, %	31.9 ^g	31.8 ^g	37.3 ^f	38.0 ^f	40.8 ^{ef}	41.4 ^{def}	45.4 ^{bcd}	46.5 ^{bc}	41.5 ^{def}	41.9 ^{def}	46.1 ^{bc}	46.9 ^{bc}	46.3 ^{bc}	48.7 ^b	48.0 ^b	58.0ª	0.9
c h ⁻¹	0.01 ^d	0.02 ^{cd}	0.02 ^{cd}	0.03 ^c	0.03°	0.01 ^d	0.04 ^b	0.05 ^b	0.02 ^{cd}	0.02 ^{cd}	0.04 ^b	0.1 ^a	0.02 ^{cd}	0.03 ^c	0.04 ^b	0.1 ^a	0.02

Table 2. Kinetics of *in situ* degradability of fallen leaves of *Quercus rugosa* treated with different levels of molasses and urea

a = fraction of dry matter lost during washing; b = fraction of dry matter slowly degraded; c = degradation rate of dry matter of fallen leaves treated with urea and molasses. SEM= standard error of the mean.

^{a, b, c, d, f, g} Mean in a row with different letter superscripts differ (P<0.05).

4. Conclusions

The OM, NDF, ADF and NFC contents in leaf litter samples diminished as the levels of urea and molasses augmented. However, CP, hemicellulose, ME, NEl and SCFA augmented as the levels of urea and molasses increased. In general, the *in situ* digestion parameters **a**, **b**, **a**+**b** and **c** of the dry matter of fallen leaves were improved as the levels of urea and molasses increased. These results confirm that the nutritive value of fallen leaves can be improved if are ensiled with urea-molasses

Corresponding Author:

Dr. Jairo Iván Aguilera Soto Unidad Académica de Medicina Veterinaria y Zootecnia Calera Campus, Universidad Autónoma de Zacatecas Zacatecas, 98560, México. E-mail: <u>aguileraivan@yahoo.com.mx</u>

References

- [1]. De la Paz POC, Campos AR, Quintanar AI, Dávalos RS. Estudio anatómico de la madera de cinco especies del genero *Quercus* (*Fagaceae*) del estado de Veracruz. Madera y Bosques 1998; 4(2): 45-65.
- [2]. Valencia AS. Diversidad del género Quercus (Fabaceae) en México. Bologial Society of Botany Mexico 2004; 75:33-53.
- [3]. Reis RA, Andrade P, Rosa B, Alcalde CR, Jobim CC. Efeito da suplementação protéica sobre o valor nutritivo da palha de aveia preta tratada com amonia. Revista Sociedad Brasileida da Zootecnia 1995; 24(2): 233-241.
- [4]. Sahoo A, Elangovan AV, Mehra U, Singh R. Catalytic supplementation of urea-molasses on nutritional performance of male Buffalo (Bubalus bubalis) calves. Asian Australasian

Journal of Animal Sciences 2004; 17(5): 621-628.

- [5]. Van Thu N, Uden P. Effect of work and ureamolasses cake supplementation on live weight and milk yield of Murrah buffalo cows. Asian Australasian Journal of Animal Sciences, 2000; 13(9): 1329-1336.
- [6]. Souza O, Dos Santos IE. Digestibilidad in vivo, balance de nitrógeno e Ingestión voluntaria en ovinos alimentados con paja de cebada tratada con urea. Archivos de Zootecnia 2002; 51: 361-371.
- [7]. Montiel MD, Elizalde JC, Santini F, Giorda L. Desactivación de taninos en grano húmedo de sorgo con polietilenglicol o urea. Archivos de Zootecnia 2012; 61: 234-241.
- [8]. Ørskov ER. The in situ technique for the estimation of forage degradability in the ruminants. In: Evaluation in Ruminant Nutrition, CAB International (eds. D.I. Givens, E.Owen, R.F.E. Axford y H.M.Ohmed) 2000:175-188.
- [9]. CONAFOR. Programa Nacional de Protección contra incendios Forestales. Comisión nacional Forestal. SEMARNAT. México 2009.
- [10]. INEGI. Prontuario de información geográfica municipal de los Estados Unidos Mexicanos. Tequila, Jalisco. Clave Geoestadística 14094 2009a.
- [11]. INEGI. Prontuario de información geográfica municipal de los Estados Unidos Mexicanos. Teúl de González Ortega. Clave Geoestadística 32047 2009b.
- [12]. INEGI. Prontuario de información geográfica municipal de los Estados Unidos Mexicanos. San Cristóbal de la Barranca, Jalisco. Clave Geoestadística 14071 2009c.
- [13]. Van Soest PJ, Robertson JB, Lewi BA. Methods for dietary fiber, neutral detergent fiber y nonstarch polysaccharides in relation to animal nutrition. Journal of Dairy Science 1991; 74: 3583-3597.
- [14]. Sniffen CJ, O'Connor JD, Van Soest PJ, Fox DG, Russell JB. A net carbohydrate and protein system for evaluating cattle diets: 2. Carbohydrate and protein availability. Journal of Animal Science 1992; 70: 3562-3577.
- [15]. Association of Official Analytical Chemists. Official Methods of Analysis, 16th ed. Arlington, VA. USA. 1995.

[16]. Abas I, Ozpinar H, Can KH, Kahraman R. Determination of the metabolizable energy (ME) and net energy lactation (NEL) contents of some feeds in the marmara region by in vitro gas technique. Turkey Journal of Veterinary Animal Science 2005; 29: 751-757.

http://www.lifesciencesite.com

- [17]. Makkar HPS. In vitro gas methods for evaluation of feeds containing phytochemicals. Animal Feed Science Technology 2005; 124: 291-302.
- [18]. Theodorou MK, Williams BA, Dhanoa MS, McAllan A, France J. A simple gas production method using a pressure transducer to determine the fermentation kinetics of ruminant feeds. Animal Feed Science and Technology 1994; 48:185-197.
- [19]. National Research Council. Committee on Nutrient Requirements of Small Ruminants. Nutrient requirements of small ruminants: sheep, goats, cervids, and new world camelids. National Academic Press. 2000;220-245.
- [20]. Menke K, Steingass H H. Estimation of the energetic feed value from chemical analysis and in vitro gas production using rumen fluid. Animal Research Development 1998; 28: 7-55.
- [21]. Nocek J, Russell JB. Protein and energy as an integrated system. Relantionship of ruminal protein and carbohydrate availability to microbial synthesis and milk production. Journal of Dairy Science 1988; 71: 2070-2107.
- [22]. Chen XB. Neway Excel-A utility for processing data of feed degradability and in vitro gas production. XBC laboratory. Rowett Research Institute 1997. Aberdeen, AB2 9SB. UK.
- [23]. Ørskov ER, McDonald LM. The estimation of protein degradability in the rumen from incubation measurement weighted according to rate of passage. Journal of Agriculture Science (Cambridge) 1979; 92: 499-503.
- [24]. SAS, SAS/STAT® User's Guide (8.1Edition). SAS Inst. Inc., Cary, NC, USA 2000.
- [25]. Araiza-Rosales E, Delgado-Licon E, Carrete-Carreón FO, Medrano-Roldán H, Solís-Soto A, Murillo-Ortiz M, Haubi-Segura C. Degradabilidad ruminal *in situ* y digestibilidad in vitro de diferentes formulaciones de ensilados de maíz-manzana

adicionados con melaza. Avances en Investigación Agropecuaria 2013; 17(2): 79-96

- [26]. Vallejo-Solís MA. Efecto del premarchitado y la adición de melaza sobre la calidad del ensilaje de diferentes follajes de árboles y arbustos tropicales. Tesis (Mag Sc). CATIE, Turrialba (Costa Rica). 1995:91-106.
- [27]. Shultz E, Shultz TA, Carnevali AA, Chicco CF. Suplementación con urea-melaza y

1/12/2022

pulitura de arroz en bovinos alimentados con pastos de pobre calidad. Agronomía Tropical 1991; 21(3): 195-204.

[28]. Pinto-Ruiz R, Hernández D, Gómez H. Cobos MA, Quiroga R, Pezo D. Árboles forrajeros de tres regiones ganaderas de Chiapas, México: usos y características nutricionales. Universidad y Ciencia. Trópico húmedo 2010; 26(1): 19-31.