# Preliminary assessment of bacteria and fungi associated with the blue swimmer crab *Portunus pelagicus* and its habitat along Thondi Coast

Beema Mahin Muhammad Iqbal, Saravanan Rajendiran and Sugumar Vasudevan\*

Department of Oceanography & Coastal Area Studies, Alagappa University, Thondi Campus, Thondi-623 409.

Tamil Nadu, India

crustacealab@gmail.com

Abstract: The increase of the contamination of seawater and sea foods due to human excreta and waste disposal has become a major threat. In the present investigation, about 12 species of bacteria and 3 species of fungi were isolated from the blue swimmer crab *Portunus pelagicus* and its habitat along the coast of Thondi, Palk Bay, Tamil Nadu, India. Morphological and biochemical characterization tests were employed to identify the microorganisms. The bacterial strains were identified as *Escherichia coli*, *Klebsiella pneumoniae*, *Proteus mirabilis*, *Shigella spp*, *Salmonella spp*, *Vibrio harveyi*, *Vibrio spp*, *Staphylococcus spp*, *Bacillus subtilis*, *Bacillus cereus*, *Aeromonas spp* and *Serratia spp*. The fungal strains were identified as *Aspergillus flavus*, *Aspergillus niger* and *Penicillium spp*. The present study will be beneficial to assess the safety of the blue swimmer crab consumption on the basis of the microorganisms associated with it. And this will pave the way for safety precautions to be taken to avoid sea food contamination.

[Beema Mahin Muhammad Iqbal, Saravanan Rajendiran and Sugumar Vasudevan. **Preliminary assessment of bacteria and fungi associated with the blue swimmer crab** *Portunus pelagicus* and its habitat along Thondi Coast. *Life Sci J* 2016;13(7):60-65]. ISSN: 1097-8135 (Print) / ISSN: 2372-613X (Online). http://www.lifesciencesite.com. 7. doi:10.7537/marslsj130716.07.

Keywords: Marine bacteria; Marine Fungi; Microbial contamination.

#### 1. Introduction

*Portunus pelagicus* (blue swimmer crab) is one of the major marine crabs used for domestic consumption as well as for export (Josileen et al., 1998). It is rich in fatty acids like Omega-3 alpha-Linoleic acid and Omega-9 Oleic acid (Samiee et al., 2012). It is a fast growing, early maturing and highly fecund species that exploit a broad ecological niche (Sumpton et al., 2003). They are found in the intertidal estuaries of Indian and Pacific Oceans and the Middle-Eastern coast of the Mediterranean Sea. In India it is widely distributed throughout West Bengal, Andhra Pradesh, Tamil Nadu, Kerala, Karnataka and Maharashtra. It contributes about 30% of the total annual marine crab landings (Samuel et al., 2004).

Microbial diversity is probably high along the coastal waters because of the combined effects of euphotic zone and proximity of the shore that supplies sediments and particles from its various marine communities (Hudson and Lester, 1994; Sutha et al., 2011). Chuma (2010) demonstrated the impact of microorganisms due to the produced water discharge in near shore shallow marine waters of Nigeria. Marine microbes were found in the surface waters, lower and abyssal depths from coastal to the offshore regions. Marine bacteria have been found in seawater, sediments and marine macro organisms (Arunkumar and Karthik, 2013). Bahamdain et al. (2015) characterized the different strains of *Bacillus* obtained from the marine habitats. The bacteria belongs to

*Vibrio* species which are potential human pathogens lives in marine and estuarine environment (Broza et al., 2007; Senderovich et al., 2010).

The occurrence of fungi in marine environment has been found in deep-sea sediments as well as near shore regions. Marine fungi are helpful in the decomposition and cycling of nutrients (Das et al., 2006). Sridhar (2009) studied the fungal diversity of Pichavaram mangroves. Generally 1 ml of seawater comprises of  $10^6$  bacteria,  $10^7$  viruses and  $10^3$  fungal colonies including mortality causing pathogens and microbes which initiate fouling on host surfaces (Kubanek et al., 2003). Fungi has been reported to survive in hyper saline regions. *Aspergillus versicolor* was isolated from the hyper saline Dead Sea water (Mbata, 2008).

Food safety has become a major concern in many countries. Food borne disease leads to serious sanitary and economic significances. In the year 2000, 1.8 million people died of food borne infections because of food and water (WHO, 2007). The food borne illness is mainly associated with human excreta and waste disposal to coastal marine environment (Dib et al., 2013).

In observance of the food safety for human consumption in mind, the present study aims to isolate the bacteria and fungi associated with the blue swimmer crab *Portunus pelagicus* and its habitat along the coast of Thondi in Palk Bay, Tamil Nadu, India.

# 2. Materials and Methods

# 2.1 Sample Collection

Sea water, sediment and crab (*Portunus pelagicus*) samples were subjected to microbial analysis. All the samples were collected from Thondi coastal area (9°45'N 79°04'E) of Palk Bay, Tamil Nadu. The samples were collected from near shore as well as offshore regions. The samples were collected in sterile plastic bags and kept in cold temperature until transported to laboratory. Due to palatability and abundant occurrence in the study area *Portunus pelagicus* was selected for the present study.

# 2.2 Isolation of Microorganisms

One gram of sediment sample and 1 ml of water sample were serially diluted with sterile sea water. The crabs were dissected aseptically. Their muscle, gill and gut samples were collected and homogenized with phosphate buffered saline (PBS, pH-7.4). The samples were serially diluted from  $10^{-1}$  to  $10^{-6}$  concentrations. The dilutions of  $10^{-4}$ ,  $10^{-5}$  and  $10^{-6}$  were spread plated on Zobell marine agar plates for bacterial isolation and on Sabouraud dextrose agar plates for fungal isolation.

The plates for bacterial isolation were incubated at 37°C for overnight and the plates for fungal isolation were incubated at 28°C for 7 days. After incubation the bacterial colonies were calculated using Total Plate Count method.

# 2.3 Identification of microorganisms

#### 2.3.1 Identification of bacteria

The colonies were picked carefully based on colony morphology which includes size, shape and colour. The isolated bacterial strains were identified as per the procedure of Bergey's manual of systematic bacteriology (Brenner et al., 2005). Morphological parameters such as Gram staining and motility tests were carried out. Motility was observed using 'hanging drop method'. The morphological parameters of fungi were studied using light microscope (Ci, Nikon, Japan) under varying magnifications.

Biochemical characterization tests like Indole, Methyl Red, Voges-Proskauer, Citrate utilization, Triple Sugar Iron agar test, Catalase, Starch hydrolysis, Protein hydrolysis, Lipid hydrolysis, Coagulase, Oxidase and Urease were performed for the identification of bacteria.

### 2.3.2 Identification of fungi

The isolated fungi were identified based on morphological characteristics of the colony as well as the cell. The identification of fungi was performed based on morphological criteria by using Lactophenol cotton blue staining (Samson et al., 1984). The fungi were identified based on morphological characteristics like nature of growth, spore colour, pigmentation, fruiting body and arrangement of spores (Raper and Fennell, 1965; Domsch et al., 1980) and http://www.doctorfungus.org/thefungi/hortaea.php.

# 3. Results

Totally 12 different species of bacteria were isolated from seawater, sediment and crab samples collected from the coast of Thondi, Palk Bay, Tamil Nadu, India. Bacteria were identified on the basis of colony morphology, cell morphology and biochemical characterization. Among the bacteria isolated nine strains such as *Escherichia coli*, *Klebsiella pneumoniae*, *Proteus mirabilis*, *Shigella spp.*, *Salmonella spp.*, *Vibrio harveyi*, *Vibrio spp.*, *Aeromonas spp.* and *Serratia spp.* were Gram negative and three strains such as *Staphylococcus spp.*, *Bacillus subtilis* and *B. cereus* were Gram positive (Table 1-3).

Table 1. Morphological characteristics of the isolated bacterial colonies

Strain No.	Form	Elevation	Margin	Colony colour
Strain-1	Circular	Raised	Entire	White
Strain-2	Circular	Convex	Entire	White
Strain-3	Circular	Raised	Entire	White
Strain-4	Circular	Raised	Entire	White
Strain-5	Circular	Convex	Entire	Red
Strain-6	Circular	Convex	Entire	White
Strain-7	Circular	Convex	Entire	White
Strain-8	Circular	Convex	Entire	Yellow
Strain-9	Irregular	Flat	Lobate	White
Strain-10	Irregular	Flat	Lobate	White
Strain-11	Circular	Raised	Irregular	White
Strain-12	Circular	Convex	Entire	Orange

Table 2. Morphological and Biochemical	characteristics of isolated bacteria
--	--------------------------------------

Tests	Strain-1	Strain-2	Strain-3	Strain-4	Strain-5	Strain-6
Morphological Tests						
Grams' Staining	-	-	-	-	-	-
Shape	Rod	Rod	Rod	Rod	Rod	Rod
Motility test	+	-	+	-	+	+
<b>Biochemical tests</b>						
Indole	+	-	-	-	-	+
Methyl Red	+	-	+	+	+	+
Voges Proskauer	-	+	-	-	-	-
Citrate	-	+	+	-	+	+
Triple sugar iron	+	+	+	+	+	+
Catalase	+	+	+	+	+	+
Starch hydrolysis	-	-	-	+	+	+
Protein hydrolysis	-	-	-	+	+	+
Lipid hydrolysis	-	-	-	-	-	+
Coagulase (Tube)	-	-	-	-	-	-
Coagulase (Slide)	-	-	-	-	-	-
Oxidase	-	-	-	-	-	+
Urease	-	+	+	-	-	+

Table 2. (continued) Morphological and Biochemical charac	cteristics of isolated bacteria
---	---------------------------------

Tests	Strain-7	Strain-8	Strain-9	Strain-10	Strain-11	Strain-12
Morphological Tests						
Grams' Staining	-	+	+	+	-	-
Shape	Rod	Cocci	Rod	Rod	Rod	Short Rod
Motility test	+	-	+	+	+	+
<b>Biochemical tests</b>						
Indole	+	+	-	-	+	-
Methyl Red	+	-	+	+	-	-
Voges Proskauer	-	+	+	+	-	+
Citrate	-	-	+	+	-	+
Triple sugar iron	+	+	+	+	-	+
Catalase	-	-	+	+	+	+
Starch hydrolysis	-	-	+	+	+	-
Protein hydrolysis	+	-	+	+	+	+
Lipid hydrolysis	+	-	-	-	+	+
Coagulase (Tube)	-	-	-	-	-	-
Coagulase (Slide)	-	-	-	-	-	-
Oxidase	+	-	-	-	+	-
Urease	-	-	-	-	-	+

Table 3 List of bacteria corresponding to strain number

S. No.	Strain Number	Bacteria
1.	Strain-1	Escherichia coli
2.	Strain-2	Klebsiella pneumoniae
3.	Strain-3	Proteus mirabilis
4.	Strain-4	Shigella spp.
5.	Strain-5	Salmonella spp.
6.	Strain-6	Vibrio harveyi
7.	Strain-7	Vibrio spp.
8.	Strain-8	Staphylococcus spp.
9.	Strain-9	Bacillus subtilis
10.	Strain-10	Bacillus cereus
11.	Strain-11	Aeromonas spp.
12.	Strain-12	Serratia spp.



Fig. 1- Aspergillus flavus

Three species of fungi were isolated from the sediment and they were identified based on the morphological characteristics such as nature of growth, spore colour, pigmentation, fruiting body and arrangement of spores as *Aspergillus flavus*, *A. niger* and *Penicillium spp*. The microphotographs of the isolated fungi are presented in Figs. 1-3.

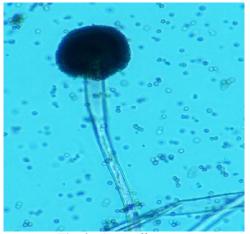


Fig. 2- Aspergillus niger



Fig. 3- Penicillium spp.

#### 4. Discussions

The microorganisms isolated in the present study could be pathogenic and involve in the health complications of the human beings. Our findings reinforce the previous reports of Immaculate et al., (2012) and Mahalaxmi et al., (2013) who reported the occurrence of *Escherichia coli* from the study site. It was reported that *E.coli* does not endure in marine environment. But in our study the existence of *E.coli* may be due to the untreated sewage disposal and improper sanitary status. The pathogenic strains of *E.coli* can cause severe infections in the intestine of human beings. The presence of *E. coli* has been implicated to be due to the occurrence of fecal contamination in the Setiu wetland, Malaysia (Najiah et al., 2010). Faghri et al., (1984) isolated the *Klebsiella* species and other enteric bacteria from the tissues of several types of crabs such as *Chionoecetes opilio*, *Cancer magister*, *Paralithodes camtschaticus* and *Cancer irroratus* collected from a contaminated area. Mahalaxmi et al., (2013) reported the incidence of *Klebsiella pneumoniae* the causative organism of pneumonia from the shells, gills and hepatopancreas of the *Portunus pelagicus*. The same organism has been isolated from our study site which implies the indication of sewage disposal into the Thondi coast.

The microorganisms like *Proteus mirabilis*, *Staphylococcus spp.*, *Vibrio spp.* and *Aeromonas spp.* isolated in our present study has been previously reported by Obiajuru and Ogbulie (2006) from Imo river basin revealing the heavy contamination of aquatic environment which affects the health of aquatic organisms. Hauxhurst et al., (1980, 1981) indicated the presence of enormous amount of bacteria from the crab tissues. Tison et al., (1982, 1984) reported the presence of *Vibrio* species which are potential human pathogens from the marine ecosystems. However, the presence of human pathogens in the marine environment leads to the cross contamination of sea foods present in that location.

Sutha et al., (2011) previously reported the bacteria like B. subtilis, Staphylococcus aureus and Serratia marscesens from Thondi coast. The results of the present study also confirms their presence in the samples studied. The bacteria belonging to the genus Bacillus has been isolated from the seawaters of Mandaicad in Tamil Nadu (Ramya et al., 2012). Bacillus has been proved to be a severe food pathogen that can cause serious illness in human beings and may even leads to death. Suvega and Arunkumar (2014) reported the presence of Aeromonas sp., E. coli, B. subtilis, B. cereus and S. aureus from the marine samples collected at different localities along the coast of South India. Vibrio spp., B. subtilis and B. cereus have been isolated from the Palk Bay sediments (Nithya and Pandian, 2010).

*Escherichia coli*, *S. typhi*, *P. aeruginosa*, *V. cholerae* and *S. dysentriae* were isolated from the muscle of two edible fishes from Royapuram waters of Chennai, India (Sujatha et al., 2011).

In our present study three fungi such as *Aspergillus flavus*, *A. niger* and *Penicillium spp.* has been reported. The salt tolerant fungi belonging to the genus *Aspergillus* and *Penicillium* were isolated from the mangroves and solar salterns of Goa (Nayak et al., 2012). Raghukumar and Raghukumar (1998) isolated the fungi *Aspergillus ustus* from the Arabian Sea and Bay of Bengal, India. *Penicillium spp.* was isolated from the deep water sediment at 4380 ft. Fungal contamination in the marine environment may leads to

the severe mortality of the marine animals in their juvenile stage (Gautschi et al., 2004).

### 5. Conclusion

In conclusion, the presence of several bacteria and fungi strains which have been reported in the present study can cause severe food borne illnesses in human beings. Previous reports have proved these strains to be severe human pathogens. The main source of these contamination is the human excreta and domestic sewage disposal into the marine environment. Hence, preventive measures like good sanitary and hygienic practices should be taken to prevent this type of contamination. The people dwelling near the Thondi coastal area should be given awareness and preventive measures should be taken to safeguard the marine ecosystem from severe microbial contamination which influence the health of humans.

#### Acknowledgement

This research work was partially financed by a project funded by the Department of Science and Technology (F.No.SR/FT/LS-137/2009), Government of India, New Delhi.

### **Corresponding Author:**

Dr. Sugumar Vasudevan, Department of Oceanography & Coastal Area Studies, Alagappa University, Thondi Campus, Thondi – 623 409. Tamil Nadu, India. Contact No.: +91 94451 39906 E. Mail: <u>crustacealab@gmail.com</u>

# References

- 1. Josileen J, Maheswarudu G, Arputharaj MR. Blue swimmer crab hatchery. In: Proceedings of the Workshop National Aquaculture Week. The Aquaculture Foundation of India, Chennai, (1998). pp. 108-110.
- Samiee K, Rustaiyan A, Far SK. Extraction and identification of natural compounds in muscle tissue of blue swimming crab (*Portunus pelagicus*) in Persian Gulf Coasts. J. Am. Sci. 2012; 8(8): 530-534.
- Sumpton W, Gaddes S, Mclennan M, Campbell M, Tonks M, Good N, Hagedoorn W, Skilleter G. Fisheries biology and assessment of the blue swimmer crab (*Portunus pelagicus*) in Queensland. Queensland Department of Primary Industries and the Fisheries Research and Development Corporation. 2003; Project No., 98/117.
- Samuel JN, Thirunavukkarasu N, Soundarapandian P, Shanmugam A, Kannupandi T. Fishery potential of commercially important portunid crabs along Parangipettai coast. In:

Proceedings of Ocean Life Food & Medicine Expo. 2004; .pp: 165-173.

- 5. Hudson DA, Lester RJG. Parasites and symbionts of wild mud crabs, *Scylla serrata* of potential significance in aquaculture. Aquaculture. 1994; 120: 183-199.
- Sutha SP, Venkatesan M, Arunkumar K. Endobiotic bacteria in some seaweeds of Thondi coastal region in Palk Bay, Tamil Nadu, India. J. Mar. biol. Ass. India. 2011; 53:251-256.
- 7. Chuma CO. Microbiological impacts of produce water discharges in near shore shallow marine waters near Chevron's Escravos tank farm, Nigeria. J. Am. Sci. 2010; 6(3): 93-101.
- Arunkumar K, Karthik R. Screening and isolation of bacteria associated with seaweeds occurring along the Coast of Thondi (Bay of Bengal, India) for Chitinolytic activity. Int J App Res. 2013; 114-18.
- 9. Bahamdain L, Fahmy F, Lari S, Aly M. Characterization of some *Bacillus* strains obtained from marine habitats using different taxonomical methods. Life Sci. J. 2015; 12(4): 58-63.
- Broza YY, Poleg YD, Lerner L, Broza M, Kashi Y. *Vibrio vulnificus* typing based on simple sequence reports: Insight into the biotype 3 group. J Clin Microbiol. 2007; 45: 2951-2959.
- 11. Senderovich Y, Izhaki I, Halpen M. Fish as reservoirs and vectors of *Vibrio cholera*. PLOS One. 2010; 5e8607-e8607.
- 12. Das S, Lyla PS, Khan SA. Marine microbial diversity and ecology: importance and future perspectives. Curr. Sci. 2006; 90:1325-1335.
- Sridhar KR. Fungal diversity of Pichavaram mangroves. Nature and Science. 2009; 7(5): 67-75.
- Kubanek J, Jensen PR, Keifer PA, Sullards MC, Collins DO, Fencial W. Targeted chemical defence against marine fungi. Proc. Natl. Acad. Sci. 2003; 100: 6916-6921.
- 15. Mbata TI. Isolation of fungi in hyper saline Dead Sea water. J Public Health. 2008; 3: 170-172.
- 16. WHO (World Health Organisation) (2007) Food safety and Foodborne Illness, Media centre. (http://www.who.int/mediacentre/factsheets/fs23 7/en/).
- 17. Dib AL, Chahed A, Elgroud R, Kabouia R., Lakhdara N, Bouaziz O, Garcia ME. Evaluation of the contamination of sea products by *Vibrio* and other bacteria in the eastern coast of Algeria. Arch Appl Sci Res. 2013; 5: 66-73.
- Brenner DJ, Krieg NR, Staley JT. Bergey's Manual of Systematic Bacteriology, Vol. 2, Springer, New York II. 2005; 491-555.

- 19. Samson RA, Hoekstra ES, Van Oorschot CAN. Introduction to food-borne fungi. Publ. Centraalbureau Voorschmmelculture baarn, Delft, Inst of Royal Netherlands Academy of Arts and sciences. 1984; 249pp.
- 20. Raper KB, Fennell DI. The genus Aspergillus, 1965 (Williams & Wilkins, Baltimore U.S.A).
- 21. Domsch KH, Gams W, Anderson TH. Compendium of Soil fungi, (1980) Vol I, IHW-Verlag, Eching.
- 22. Immaculate J, Sinduja P, Jamila P. Biochemical and microbial qualities of *Sardinella fimbriata* sun dried in different methods. International Food Research Journal 19 2012; 1699- 1703 Journal homepage: <u>http://www.ifrj.upm.edu.my</u>.
- Mahalaxmi B, Revathy K, Raghunathan C, Anjalai K, Subashini A. Distribution of microbial population associated with crabs from Ennore seacoast Bay of Bengal north east coast of India. Int. J. Curr. Microbiol. Appl. Sci. 2013; 2: 290-305.
- 24. Najiah M, Nadirah M, Sakri I, Harrison FS. Bacteria associated with wild mud crab (*Scylla serrata*) from Setiu Wetland, Malaysia with emphasis on antibiotic resistances. Pak J Biol Sci. 2010; 13: 293-297.
- 25. Faghri MA, Pennington CL, Cronholm LS, Atllas RM. Bacteria associated with crabs from cold waters with emphasis on the occurrence of potential human pathogens. Appl Environ Microbiol. 1984; 47: 1054-1061.
- 26. Obiajuru IOC, Ogbulie JN. Bacteriological quality of some fishes and crab from rivers within Imo river basin. J Aquat Sci. 2006; 21: 9-14.
- 27. Hauxhurst JD, Krichevsky MI, Atlas RM. Numerical taxonomy of bacteria from the Gulf of Alaska. J Gen Microbiol. 1980; 120: 131-148.
- Hauxhurst JD, Kaneko T, Atlas RM. Characteristics of bacterial communities in the Gulf of Alaska. Microbial Ecol. 1981; 7: 167-182.

- 29. Tison DL, Nishibuchi M, Greenwood JD, Seidler RJ. *Vibrio vulnificus* biogroup 2: New biogroup pathogenic for eels. Appl Environ Microbiol. 1982; 44: 640-646.
- Tison DL, Greenwood J, Nishibuchi M, Seidler RJ. Molecular taxonomy of Lactose-fermenting Vibrio. In: Vibrios in the environment, Colwell, R. (Ed.). John Wiley and Sons Inc., USA. 1984; 217-237.
- Ramya SR, Asha KRT, Baskaran N. Isolation and Characterization of bacterial strains from seawater of Mandaicad in Tamil Nadu, India. Int J Pharmaceut Biol Arch. 2012; 3: 1527-1532.
- Suvega T, Arunkumar K. Antimicrobial activity of bacteria associated with seaweeds against plant pathogens on par with bacteria found in seawater and sediments. Br Microbiol Res J. 2014; 4: 841-855.
- Nithya C, Pandian SK. Isolation of heterotrophic bacteria from Palk Bay sediments showing heavy metal tolerance and antibiotic production. Microbiol Res. 2010; 165: 578-593.
- 34. Sujatha K, Senthilkumaar P, Sangeetha S, Gopalakrishnan MD. Isolation of human pathogenic bacteria in two edible fishes, *Priacanthus hamrur* and *Megalaspis cordyla* at Royapuram waters of Chennai, India. Indian J Sci Tech. 2011; 4: 539-541.
- 35. Nayak SS, Gonsalves V, Nazareth SW. Isolation and salt tolerance of halophilic fungi from mangroves and solar salterns in Goa-India. Indian J Geo-Mar Sci. 2012; 41: 164-172.
- Raghukumar C, Raghukumar S. Barotolerance of fungi isolated from deep-sea sediments of the Indian Ocean. Aquat. Microb. Ecol. 1998; 15: 153-163.
- Gautschi JT, Amagata T, Amagata A, Valeriote FA, Mooberry SL, Crews P. Expanding the strategies in natural product studies of marinederived fungi: A chemical investigation of Penicillium obtained from deep water sediment. J. Nat. Prod. 2004; 67: 362-367.

7/18/2016