Modification of the glassy carbon electrode with Multi-walled carbon nanotube/1, 4-dihydroxy Anthraquinone/chitosan As a potentiometric pH sensor

Meisam Asghari

Faculty of Chemistry, University of Sistan & Baloochestan, Iran. Email: mei_asghari_chem@yahoo.com

Abstract: In this study, the electrochemical behavior of 1, 4-dihydroxy anthraquinone (Q) is investigated with cyclic voltammetry (CV) technique at glassy carbon electrode (GCE) in different pH. The results have been shown one redox wave in aqueous solution and the redox wave shifted to negative potential with increasing pH. The half potential (E $_{1/2}$) for redox wave was linear in the pH range form 1.5 to 7.0. A modified glassy carbon electrode (GCE) is prepared using an appropriate mixture of Q/multi-walled carbon nanotubes/Chitosan. The electrochemical behaviors of sensor were studied in different pH. The half potential (E $_{1/2}$) of the redox wave was shifted to negative potential with increasing pH. It is found the E $_{1/2}$ for redox wave was linear over a range of 11 pH units in pH range between 1.0 and 12.0 the values of correlation coefficient and RSD are 0.997 and 1.99% respectively which is so better than previous method.

[Meisam Asghari. Modification of the glassy carbon electrode with Multi-walled carbon nanotube/1, 4-dihydroxy Anthraquinone/chitosan As a potentiometric pH sensor. *Life Sci J* 2013;10(10s):16-18]. (ISSN:1097-8135). http://www.lifesciencesite.com. 3

Key words: multi-walled carbon nanotubes, Chitosan, pH sensor, 1, 4-dihydroxy anthraquinone

1. Introduction

Chemical sensors provide direct, real-time information on the contents of certain chemical substance(s) present in their environment [1]. Their practical importance continuously increases as they not only offer an advantageous alternative to time- and cost demanding laboratory analyses but, primarily, feed necessary input information to a great variety of automatic devices, regulating mechanisms and robots; therefore, humans often cease to be direct users of the information provided and can concentrate on the designing and operating larger scientific technological systems. Sensor signals are handled by electronic circuits and it is advantageous, for financial, spatial and operational reasons, to integrate the sensor and its electronic circuitry into a single block; in this way it is further possible to construct integrated systems with sensor arrays and systems with some degree of artificial intelligence (smart sensors).

A significant group of chemical sensors is based on electrochemical principles [1, 2]. The classical electrochemical sensors with liquid electrolytes are usually rather bulky and awkward; moreover, drying or leakage of the electrolyte may corrode various parts of the device and thus limit its lifetime and impair its function. Therefore, the progress in the field is directed towards solid-state sensors, i.e., those that contain no macroscopic liquid phase. There exist several ways for preparation of solid-state electrochemical sensors and one of the important approaches is the replacement of a liquid electrolyte by a solid polymer exhibiting an ionic conductivity, called a solid polymer electrolyte (spe). On the other hand pH

is one of the most common laboratory measurements because so many chemical, biological processes and practical applications such as clinical analysis, environmental analysis, process control and rate of chemical and biochemical reactions are dependent on pH [3, 4]. In tradition, the measuring methods for pH values fall roughly into four categories: indicator reagents, pH test strips, glass electrode and metal electrode methods (hydrogen electrode, quinhydrone electrode and antimony electrode method). In past, the hydroquinone electrode used for determination of pH with potentiometric method but today this method is less used because not reproducible [5]. Obviously MWCNT as a magical material, because their uniqueness and have received excellent attention for the preparation of electrochemical sensor, as it was extensively reviewed. [6, 11]

2. Experimental

2.1. Apparatus

All the electrochemical measurements were carried out with an Electro analyzer.

The three-electrode system consisted of a modified GCE, a SCE reference electrode, and a platinum auxiliary electrode. All potentials were referred to the SCE.

2.2. Reagents

The chitosan solution (8%) was prepared by dissolving 0. 8 g chitosan in 10 ml acetic acid. CNTs functionalized by sonification treatment in a mixture of sulfuric acid and nitric acid (3:1) for 6 h at room temperature. Functionalized MCNTs (fMCNTs) were

washed with dowble distilled water (DDW) and separated by centrifugation. [12].

1,4-Diydroxy anthra quinone(Q), Acetonitrile (AN). All experiments were carried out at room temperature (about 15 °C).

2.3. Preparation for MWCNT-chitosananthraquione modified electrode

First make a solution of Q (0.05mg) in 2.5ml acetonitril and then added 25mg of MWCNT and agitation with ultrasonic to give a black suspension, now 0.5ml of this suspension mixed with 0.1ml of chitosan solution (0.5%).

Before being modified, the GCE was polished with 0.3 and 0.05 μm aluminum slurry, rinsed thoroughly with DDW, then ultrasonically cleaned with $H_2SO_4(1M)$ for 5 min, finally rinsed with DDW, and dried under an infrared lamp. After the GCE was cooled, it was smeared evenly with 6 μL of a MWCNT–chitosan-anthraquione (0.5%) solution by a micro-syringe, and then dried under an infrared lamp for 10 min. After cooling, the MCQ/GCE could be used.

2.4. Experimental procedures

A phosphate buffer solution (PB) 0.3 M (pH=1-12) was used as the supporting electrolyte. The solution was purged with nitrogen for 2 minutes. In the differential pulse Volta metric (DPV) study, the electrode was scanned between 0 and -1V.

3. Results and discussion

3.1. Electrochemical Behavior of O

the electrochemical behavior of 1, 4-dihydroxy anthraquinone (Q) is investigated with cyclic voltammetry (CV) technique at glassy carbon electrode (GCE) in different pH. The results have been shown (Figure 1) one redox wave in aqueous solution and the redox wave shifted to negative potential with increasing pH. The catodic potential ($E_{\rm pc}$) for redox wave was linear in the pH range form 1.5 to 7.0 with a correlation coefficient of r=0.97.

3.2. Electrochemical Behavior of OMC/GCE

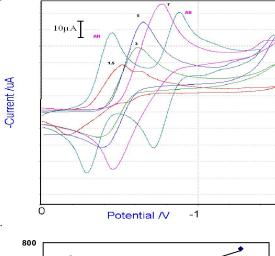
The CV former component of elecrode shown in fig.2 at the QMC/GCE the CV demonstrated two reversible waves with half peaks potential $(E_{\rm 1/2})$ respectively at -0.032and -0.327 V.

The behavior of Q at a bare GCE and QMC/GCE was studied by cyclic voltammetry.

As shown in fig.2 the values of i_{pa} , i_{pc} for QMC/GCE is more than of this values for Q/ GCE respectively because chitosan promote electron transfer in the matrix [13-15].

The electrochemical behaviors of sensor were studied in different pH in DPV mode. The half potential ($E_{1/2}$) of the redox wave was shifted to negative potential with increasing pH. It is found the $E_{1/2}$ for redox wave was linear over a range of 11 pH units in pH range between 1.0 and 12.0 with a correlation coefficient of 0.997, which is so better than

previous method[16]. Because this electrode is stable at this range of pH and on the other hand Better performance for detection of pH on MWCNT-GCE was attributed to the MWCNTs potential porous leading to the larger electroactive surface area and thus greater response of pH [17,18]. This result are given in fig.3.



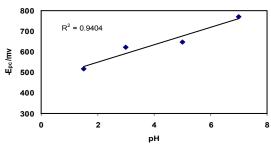


Figure 1. Cyclic voltammogram for 5.0×10^{-3} mol L⁻¹ Q in mixture (3:1) acetonitril and PB solution (PH=1.5-7) + 0.1 M TBAP on GCE in the, Scan rate: 100 mVs-1. Inset, plot of $E_{1/2}$ vs. pH values

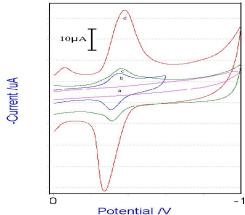


Fig 2: The CV former component of electrode in 0.3M PB (PH=1.5), Scan rate=100mv/sec,a)modified GCE with chitosan (C/ GCE) b) Q/ GCE c) QC/ GCE d) QMC/ GCE

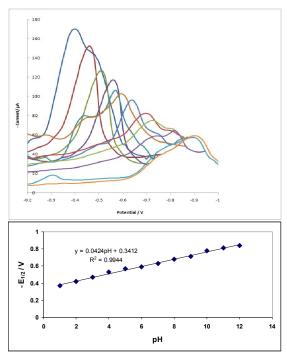


fig.3 Diffrential pulse voltammograms of QMC/GCE, at scan rate 100 mVs-1 with increasing pH values (from 1 to 12) 1.0, 2.0, 3.0, 4.0, 5.0,6.0,7.0,8.0,9.0,10.0,11.0,12.0 in phosphate buffer solution. Inset, plot of $E_{1/2}$ vs. pH values.

3.3. Effect of the amount of Q–MWCNT modifier at the QMC/GCE $\,$

The amount of modifier on the surface of electrode affected the i_{pa} and i_{pc} . Fig. 4 showed that the i_{pa} , i_{pc} enhanced when the amount of MWCNT increased, which probably because of the presence of electro-reactive sites which increased with the amount of MWCNTs[17.18].

When the concentration of MWCNTs in the modifier surpassed 0.5ml, the i_{pa} , i_{pc} of pH fell. These because the film on the electrode surface was too thick, which prevented the electron transfer between Hydrogen ions and electrode.

In this paper, 0.5ml MWCNTs in the Q-chitosan–MWCNT modifier was used as optimum amount for voltammetric determination of pH.

Conclusions

We describe a highly sensitve sensor for monitoring of pH in wide range, although in value of more pH the quinine compounds are not stable at the surface of electrode but as mentioned this modified electrode stable and reproducibile. in different pH and scan rates.on the other hand the electrochemical behavior of Q investigated in different medium and pH at BGC and M-GC. Obviously we found that the

chitosan is good compound for immobilization of Q at the surface of electrode.

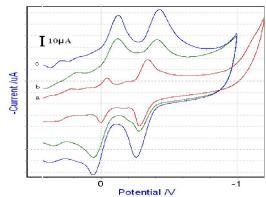


Fig. 4 The cyclic voltammogram of QMC/GCE preparated with ratios contains a) 0.1 ml of mix (MWCNT, Q) b) 0.3 ml of mix (MWCNT, Q), c)0.5 ml of mix (MWCNT, Q) with 0.1 ml chitosan in 0.3M PB (PH=1.5), Scan rate=100mv/sec

References

- W. GoÈ pel, J. Hesse, J.N. Zemel (Eds.), Sensors. A Comprehensive Survey, vols. 2 and 3, VCH, Weinheim, 1989.
- J. Janata, M. Josowicz, D.M. DeVaney, Anal. Chem. 66 (1994) 207R.
- E.Lindner, T.Zwicki, E.Bakker, B.T.T. Lan, K. Toth, E. Pretsch, Anal. Chem. 70(1998) 1176.
- H. Hisamato, M. Tsubuku, T. Enomoto, K. Watanabe, H. Kawaguchi, Y. Koike, K. Suzuki, Anal. Chem. 68 (1996)3871.
- Z.Galback, H.V. Langenhov, G, Galbacs, Talanta, 66 (2005) 809.
- M. Trojanowicz, M. Szewczynska, TrAC-Trend. Anal. Chem. 24(2005) 92.
- 7. J. Wang, Electroanalysis 17 (2005) 7.
- 8. N. Li, J. Wang, M. Li, Rev. Anal. Chem. 22 (2003) 19.
- 9. C.E. Banks, R.G. Compton, Analyst 131 (2006) 15.
- 10. [J. Wang, Analyst 130 (2005) 421.
- M. Valcarcel, B.M. Simonet, S. Cardenas, E.B. Suarez, Anal. Bioanal. Chem. 382(2005) 1783.
- S.Hoon Baek, B. Kim, K.Do Suh, Colloids and Surfaces A: Physicochem. Eng. Aspects 316 (2008) 292–296.
- 13. Hang M, Gorski W. J Am Chem Soc (2005) 127:2058–9.
- 14. Zhang M, Gorski W.Anal Chem (2005)77:3960-5.
- Zhang M, Smith A, Gorski W. Anal Chem (2004) 76:5045–50.
- S. Rosini, E. Siebert, Electrochimica Acta 49 (2004) 525–536
- 17. C.E. Banks, T.J. Davis, G.G. Wildgoose, R.G. Compton, Chem.Commun. 7 (2005) 829.
- C.E. Banks, R.R. Moore, T.J. Davis, R.G. Compton, Chem. Commun. 16 (2004) 1804.

9/28/2013