

Lipoic Acid Attenuates Cholestasis Induced Cerebral Injury in Rats

Mohammad Mehdi Hosseinian Zakaria¹, Babak Hajipour^{*2}, Mohammad Taghizadieh³, Ghafour Mousavi⁴, Ali Khodadadi⁵

¹Department of Neurology, Medical Faculty, Tabriz Branch, Islamic Azad University, Tabriz, Iran

²Department of Surgery, Urmia University of Medical Sciences, Urmia, Iran

³Department of Pathology, Medical Faculty, Tabriz Branch, Islamic Azad University, Tabriz, Iran

⁴Department of Surgery, Veterinary Faculty, Tabriz Branch, Islamic Azad University, Tabriz, Iran

⁵Department of Clinical Pathology, Veterinary Faculty, Tabriz Branch, Islamic Azad University, Tabriz, Iran

* **Corresponding Author:** Hajipourb@yahoo.com

Abstract: Cholestasis is characterized by an abnormal accumulation of bile acids, which is caused by defectiveness in the process of bile acid transport. It is believed that oxidative stress is a likely mediator for cholestatic damage and antioxidant therapy is a recommended therapeutic strategy. The aim of this study was to evaluate protective effect of alpha lipoic acid as an anti oxidant agent on cerebral injury after bile duct ligation in rats. forty five adult male wistar rats were randomly assigned to three groups each containing fifteen rats as follows: sham operation (SO) (control), bile duct ligation (BDL), and BDL+LA (25mg/kg). After fourteen days cerebral tissue sampled for pathologic and biochemical studies. Levels of SOD and GPx antioxidant enzymes were higher in BDL+LA group comparing to BDL group significantly, histologic damage and MDA levels were higher in BDL group comparing to BDL+LA group significantly (P<0.05). In our study LA treatment in BDL rats improved cellular SOD and GPx levels and reduced MDA levels in BDL+LA group comparing to BDL group. The findings of our present study showed that LA, with its potent free radical scavenging and antioxidant properties, seems to be a highly promising agent in protecting cerebral tissue against oxidative damage.

[Mohammad Mehdi Hosseinian Zakaria, Babak Hajipour, Mohammad Taghizadieh, Ghafour Mousavi, Ali Khodadadi. **Lipoic Acid Attenuates Cholestasis Induced Cerebral Injury in Rats.** *Life Sci J* 2013;10(5s):539-545] (ISSN:1097-8135). <http://www.lifesciencesite.com>. 95

Keywords: Lipoic Acid- Brain-Cholestasis-Rat.

1. Introduction

Cholestasis is characterized by an abnormal accumulation of bile acids, which is caused by defectiveness in the process of bile acid transport. Bile acids are the major products of cholesterol metabolism in the liver, and act as physiological detergents that facilitate absorption, transport, disruption of lipid-soluble fats and vitamins; furthermore it also aids in the excretion of lipids. Retention and accumulation of toxic, hydrophilic bile salts stimulates the production of proinflammatory cytokines and enhances apoptosis which leads to tissue damage (Trauner et al., 1998; Miyoshi et al., 1999). Apoptosis is an integral part of many biological processes, including embryonic development, metamorphosis, hormone-dependent atrophy, and in chemical-induced cell death (Allen et al., 1997; Patel et al., 1994). Hepatic encephalopathy is a well described clinical entity associated with obstructive jaundice and liver failure. The pathophysiological cascade responsible for central nervous system dysfunction under conditions of hepatopathy is not fully elucidated. It is considered to implicate many factors, ranging from ammonia and manganese neurotoxicity (Seyan et al., 2010) to

inflammatory cytokines (Seyan et al., 2010) and oxidative stress acting both independently and synergistically. Many cirrhotic patients, up to 50% to 70%, develop hepatic encephalopathy (Quero JC et al., 1996). A neuropsychiatric syndrome characterized by alterations of intellectual function, personality, consciousness and motor coordination (Erceg et al., 2004).

The bile acid concentrations increase in rats after BDL and induce lipid peroxidation, which is probably related to the stimulation of phagocytic activity in polymorphonuclear phagocytes and inflammatory cells (Tomioaka et al., 2000; Rivera-Mancía et al., 2009). Therefore, it is believed that oxidative stress is a likely mediator for cholestatic damage and antioxidant therapy is a recommended therapeutic strategy. Alpha-lipoic acid (LA) or thioctic acid (chemical name: 1,2 dithiolane-3-valeric acid or 6,8-dithio-octanoic acid) is a natural dithiol compound which is known as a co-factor in the α -ketoacid dehydrogenase mitochondrial complex and for its complex antioxidant properties (Moini et al., 2002; Biewenga et al., 1997; Bilaska et al., 2005). Initially, α -lipoic (LA) was obtained from livers and it has been found naturally in many plants and

animals (Reed et al., 1951). It is absorbed from the diet, biological membranes, and is then taken up by cells and tissues (Packer et al., 1996). LA is easily absorbed and converted into the reduced form of dihydrolipoic acid in a variety of cellular tissues (Packer et al., 1998). Both act as an antioxidant in different environments and mutually form a redox couple. Alpha-lipoic acid, which has been shown to be effective in both the somatic and the autonomic neuropathies in diabetes, normalizes the endoneural bloodflow (Nagamatsu et al., 1995), reduces oxidative stress (Low et al., 1997; Nickander et al., 1996), and improves vascular dysfunction (Morcos et al., 2001; Xie et al., 2012; Vasdev et al., 2011). The aim of this study was to evaluate protective effect of alpha lipoic acid as an anti oxidant agent on cerebral injury after bile duct ligation in rats.

2. Materials and methods

2.1. Animals

Male wistar rats were obtained from laboratory animals care center of Tabriz University of Medical Sciences (Tabriz, Iran). They were allowed free access to a commercial standard diet and water ad libitum. Rats were randomly assigned to three groups, each containing fifteen rats as follows: sham operation, (control), BDL, and BDL+LA. Sham-operated rats served as controls. Except in this group, biliary canals were ligated. Rats were fasted for 12 h before the operation, but were given water.

2.2. Surgery protocol

The animals were anesthetized by intramuscular injection of 50 mg/kg ketamine hydrochloride and 10 mg/kg xylazine. Midline laparotomy was performed under sterile conditions. In sham group, the common bile duct (CBD) was freed from the surrounding soft tissue, and was manipulated without ligation and transaction. In BDL and BDL+LA groups, the CBDs of the rats were identified, double ligated with 5-0 silk, and divided between the ligatures. BDL+LA group was administered by LA 25mg/kg subcutaneously for 14 days (Mythili et al., 2007). The animals were sacrificed on 14th postoperative day with high-dose diethyl ether inhalation. Subsequently, the cerebral tissue was obtained.

2.3. Light Microscopy Analyses

After decapitation, the left hemispheres of the brains were stored in the 10% formaldehyde overnight at 4°C. The samples were then fixed in a 10% buffered formalin solution for 7 days. The left hippocampal regions were obtained from coronal sections of the frontal planes. Formaline-fixed, paraffin-embedded sections (4- μ m thickness) were stained with hematoxylin eosin and cresyl violet.

Intact hippocampal CA1 pyramidal neurons were semiquantitatively counted in three consecutive 789 μ m² areas outlined with a counting Gundersen's frame (Gundersen et al., 1988) under 40x magnifications, and in three consecutive hippocampus sections. The histologist was blinded to the animal groups, and the procedure was conducted in a blinded fashion (Onem et al., 2006).

2.4. Assay of antioxidant enzymes

The hippocampus was excised and frozen in liquid nitrogen and stored at -80°C until further preparation. In order to measure anti-oxidant enzyme activity, the samples were homogenized in 1.15% KCL solution. Superoxide dismutase (SOD) activity in tissue was determined by using xanthine and xanthine oxidase to generate superoxide radicals which then react with 2-(4-iodophenyl)-3-(4-nitrophenol)-5-henyltetrazolium chloride to form a red formazan dye. The SOD activity was then measured by the degree of inhibition of this reaction (Ransod, Randox Laboratories Ltd. United Kingdom). Results obtained as SOD Unit/mg protein (Paoletti et al., 1986).

Glutathione peroxidase (GPx) activity was measured using the method described by Paglia and Valentine. GPx catalyses the oxidation of glutathione by cumene hydroperoxide. In the presence of glutathione reductase and NADPH, the oxidised glutathione is immediately converted to reduced form with a concomitant oxidation of NADPH to NADP⁺. The decrease in absorbance at 340 nm is measured (Ransod, Randox Laboratories Ltd. United Kingdom). Results obtained as GPx Unit/mg protein (Paglia et al., 1967).

2.5. Tissue MDA level

Tissue malondialdehyde was determined by the method of Uchiyama and Mihara (Mihara et al., 1983) 3-mL aliquot of 1% phosphoric acid and 1 mL of 0.6% thiobarbituric acid solution were added to 0.5 mL of 10% tissue homogenate. The mixture was heated in boiling water for 45 minutes. After cooling, the color was extracted into 4 mL of n-butanol. The absorbance was measured in a spectrophotometer at 532 nm ($\epsilon = .56 \times 10^5 \text{ mol/L}^{-1} \text{ cm}^{-1}$). The amounts of lipid peroxides calculated as thiobarbituric acid reactive substances of lipid peroxidation were expressed as nMol/ml (Kirimlioglu et al., 2008).

2.6. Statistical analysis

Data were expressed as means \pm SD. Differences among various groups were tested for statistical significance using the one-way ANOVA test and Tukeys post test. A P value of less than 0.05

denoted the presence of a statistically significant difference.

3. Results

3.1. SOD and GPx level

Levels of SOD and GPx antioxidant enzymes were decreased in hippocampus of the groups subjected to bile duct ligation, but it was less severe in LA treated group. SOD and GPx levels in BDL+LA group were higher than BDL group significantly ($P < 0.05$, Table 1).

Table 1: Glutathione peroxidase (GPx), Superoxide dismutase (SOD) and Malondialdehyde (MDA) levels in hippocampus tissue of rats after bile duct ligation

	MDA (nMol/ml)	GPx (Unit/mg protein)	SOD (Unit/mg protein)
Sham	0.60±0.31	2.86±0.28	2.66±0.19
BDL	1.88±0.27	1.88±0.22	1.63±0.12
BDL+LA	1.07±0.27	2.46±0.32	2.29±0.23

Note. The values are shown as a mean ±SD for rats in each group and difference of ($P < 0.05$) considered significant.

3.2. MDA level

MDA level as an index of lipid peroxidation increased significantly in hippocampus tissue after bile duct ligation. MDA level was lower in BDL+LA group comparing to BDL group significantly ($P < 0.05$) and it was lower in sham group comparing to BDL+LA group significantly ($P < 0.05$, Table 1).

3.3. Histopathology

Studying the histologic samples by light microscopy showed that, the pyramidal neurons in the subfield of the hippocampus were completely normal in appearance in sham group. Number of neurons were higher in the CA1 subfield in the BDL+LA group comparing to BDL group significantly ($P < 0.05$, Table 2).

Table 2: Number of surviving CA1 cells in hippocampus of rats

	Sham	BDL	BDL+LA
CA 1 cells number	198.50±12.44	167.90±10.40	180.70±10.37

Note. Number of CA1 neuron were higher in BDL+LA group than BDL group significantly ($P < 0.05$)

4. Discussion

Cholestasis is encountered in a variety of clinical disorders. It is the main feature of a number of chronic progressive liver diseases, including

primary biliary cirrhosis, primary sclerosing cholangitis, allograft rejection, iatrogenic obstruction of bile ducts, and biliary atresia. Cholestasis is now recognized as a disorder characterized by liver oxidants overload (Huang et al., 2003; Portincasa et al., 2007; Sastre et al., 2007). Furthermore, the oxidative stress in cholestatic liver disease is a systemic phenomenon (Ljubuncic et al., 2000; Assimakopoulos et al., 2006), probably encompassing all tissues and organs, even those separated by the blood-brain barrier (Chroni et al., 2006). Similarly, oxidative stress plays an important role in the pathogenesis of toxic tissue injury (Feher et al., 1998).

To reduce the detrimental effects of ROS, besides diminishing its production, organisms have developed their own antioxidant mechanisms including low-molecular-weight antioxidant molecules, i.e., glutathione, melatonin and various antioxidant enzymes, such as SOD and GPx and glutathione reductase. These enzymes activities are higher in the liver than in other tissues (Yuan et al., 2005). Superoxide dismutase (SOD), an oxygen radical scavenger, which converts the superoxide anion radical present in the upper stream of reactive oxygen metabolism cascade, will afford protection from cell damage (Minor et al., 1993). SOD catalyses the dismutation of the superoxide anion (O_2^-) into H_2O_2 ; GSH-Px is a selenoprotein, which reduces lipidic or nonlipidic hydroperoxides as well as H_2O_2 while oxidizing GSH (Michiels et al., 1994). In our study, we found that cholestasis impaired these enzymes activities, as indicated by the markedly lower activities compared with sham group. LA administration maintained the activities of these enzymes significantly comparing to control group ($P < 0.05$). LA is an antioxidant substance that can react at many levels: (1) it neutralizes free radicals formed by direct radical scavenging (hydroxyl radical: $HO\bullet$), hypochlorous acid and singlet oxygen), (2) it regenerates endogenous antioxidants (GSH, vitamin C, and vitamin E) from their oxidized forms, and (3) it complexes transitional metals (especially iron and copper which are involved in $HO\bullet$ synthesis) (Cakatay et al., 2006). Concerning the clinical ways, dietary antioxidants have attracted attention as preventive and therapeutic agents (Dhalla et al., 2000; Buonocore et al., 2007; Marchioli et al., 1999).

MDA is a secondary product of oxidative stress formed during lipid peroxidation and it is released as a result of the toxic effect of reactive oxygen species in rats after bile duct ligation (Orellana et al., 2000). Increased concentrations of MDA reflect the level of lipid peroxidation in tissues and it is considered as a marker of tissue injury (Draper et al., 1990). There

are several reports indicating that levels of MDA increases after bile duct ligation in rats (Canturk et al., 1998; Karaman et al., 2003). Our results are in agreement with previous works reporting high levels of MDA. In the present study, levels of MDA in the LA-treated rats were significantly lower than in the BDL group. Although tissue MDA levels were clearly decreased by LA, its exact mechanism is not known. Reductions in MDA levels in the LA-treated rats may be due to its antioxidant and free-radical scavenging effect. By protecting cell membranes, LA probably reduces the deleterious effects of oxidative stress in living cells (Shaafi et al., 2011; Ying et al., 2010).

Huang et al., (2009) reported that, melatonin treatment decreased liver and systemic oxidative stress, increased liver antioxidant activity, and improved spatial memory in developing rat with BDL-induced cholestasis. They showed that BDL-induced cholestasis in developing rats had worse spatial memory and increased liver and systemic oxidative stress as compared with jaundice-free rats; 2) melatonin treatment, in a dose-dependent manner, decreased liver and systemic oxidative stress, increased liver antioxidant activity, and improved spatial memory in developing rat with BDL-induced cholestasis. The underlying mechanisms of increased systemic oxidative stress during cholestasis may be due to the retention of toxic bile acids, which stimulate the generation of reactive oxygen species (ROS) in hepatocytes and live mitochondria (Sastre et al., 2007), and consequently hepatocellular necrosis and apoptosis. The increased ROS may cause target organs damage (e.g., brain, heart, and kidney) via systemic circulation (Ljubuncic et al., 2000; Tokaç et al., 2013; Liu et al., 2012). Huang et al., (2010) reported that cholestatic rat had a poorer performance in acquisition memory when compared with jaundiced-free rat. Ammonia exerts a deleterious effect on cerebral function and is considered to play an important role in the pathogenesis of hepatic encephalopathy (Lockwood et al., 2004). In this regard, Jover et al., (2006) added hyperammonia diet in BDL rat to simulate hepatic encephalopathy that occurs in humans. They suggest that, systemic oxidative stress, instead of ammonia, plays a role in the cognitive deficit in young rat with BDL-induced cholestasis. ROS are involved in several diseases, including ischemic injury, Alzheimer's disease, Parkinson's disease, and Down's syndrome all of which affect cognitive processes (Sayre et al., 2001; Perry et al., 2002; Butterfield et al., 2007). ROS can cause disruption of calcium homeostasis, membrane damage, and cell death (Keller et al., 1998), and has a detrimental effect on several key enzymes involved in glutamate and glucose transport (Keller et al.,

1998; Lauderback et al., 2001); all of the above-mentioned biologic effects can result in cognitive deficit. Although, we could not detect either MDA or GSH/GSSG differences in brain cortex or hippocampus, we postulate that other players in the oxidants/antioxidants system might play a role. Alternatively, other brain regions that are involved in spatial memory might account for the cholestasis induced spatial dysfunction in our rats. Generation of ROS contributes to endothelial and cellular dysfunction, resulting in increased BBB permeability and cerebral edema. Besides the breakdown of BBB barrier, the failure of the Na/K pumps, and the altered electrolyte balance of the cell, may also contribute to brain edema and pathological changes in the cellular function (De Vries et al., 1997). With respect to brain pathology, in experimental animals, histologic changes consisting of atrophy, pyknosis, and neuronophagia were observed after 7 days of obstructive jaundice at the basal ganglia, putamen, and red nucleus, whereas after 13 days these changes were spread to the thalamus substantia nigra and cortex of the canine brain (Furukawa et al., 1991). The current study revealed that markers of oxidative stress, which could eventually lead to structural damage, were present as early as 5 days after the bile duct ligation. In our study 14 days after BDL, number of neurons in hippocampus was reduced significantly, while treatment of LA attenuated the reduction of neurons in hippocampus comparing to control group significantly. It has been proposed that antioxidant therapy may be useful for preventing the deleterious effect of oxidative stress on the nervous system during BDL (Chroni et al., 2006).

5. Conclusion

In our study LA treatment in BDL rats improved cellular SOD and GPx levels and reduced MDA levels in BDL+LA group comparing to BDL group. The findings of our present study demonstrate that LA, with its potent free radical scavenging and antioxidant properties, seems to be a highly promising agent in protecting cerebral tissue against oxidative damage.

Acknowledgment:

This research is supported by a grant from Tabriz branch, Islamic Azad University, Tabriz, Iran.

Reference:

1. Allen RT, Hunter WJ, Agrawal DK. Morphological and biochemical characterization and analysis of apoptosis. *J Pharm Toxicol Methods* 1997;37:215-228.
2. Assimakopoulos SF, Thomopoulos KC, Patsoukis N, Georgiou CD, Scopa CD, Nikolopoulou VN,

- Vagianos C. Evidence for intestinal oxidative stress in patients with obstructive jaundice. *Eur J Clin Invest* 2006;36:181–187.
3. Biewenga GP, Haenen GR, Bast A. The pharmacology of the antioxidant lipoic acid. *Gen Pharmacol* 1997;29:315–331.
 4. Biliska A, Wlodek L. Lipoic acid-the drug of the future? *Pharmacol Rep* 2005;57:570–577.
 5. Buonocore G, Groenendaal F. Anti-oxidant strategies. *Semin Fetal Neonatal Med* 2007;12:287–295.
 6. Butterfield DA, Reed T, Newman SF, Sultana R. Roles of amyloid beta-peptide-associated oxidative stress and brain protein modifications in the pathogenesis of Alzheimer's disease and mild cognitive impairment. *Free Radic Biol Med* 2007;43:658–677.
 7. Cakatay U. Pro-oxidant actions of alpha-lipoic acid and dihydrolipoic acid. *Med Hypotheses* 2006;66:110–117.
 8. Canturk NZ, Canturk Z, Utkan NZ, Yenisey C, Ozbilim G, Yildirim C, Yalman Y. Cytoprotective effects of alpha tocopherol against liver injury induced by extrahepatic biliary obstruction. *East Afr Med* 1998;8:75-77.
 9. Chroni E, Patsoukis N, Karageorgos N, Konstantinou D, Georgiou C. Brain oxidative stress induced by obstructive jaundice in rats. *J Neuropathol Exp Neurol* 2006;65:193–198.
 10. Chroni E, Patsoukis N, Karageorgos N, Konstantinou D, Georgiou G. Brain oxidative stress induced by obstructive jaundice in rats. *J Neuropathol Exp Neurol* 2006;65:193–198.
 11. De Vries HE, Kuiper J, De Boer AG, et al. The bloodbrain barrier in neuroinflammatory diseases. *Pharmacol Rev* 1997;49:143-55.
 12. Dhalla NS, Elmoselhi AB, Hata T et al. Status of myocardial antioxidants in ischemia-reperfusion injury. *Cardiovasc Res* 2000;47:446–456.
 13. Draper HH, Hadley M. Malondialdehyde determination as index of lipid peroxidation. *Methods Enzymol* 1990;186:421.
 14. Erceg S, Felipe V. Alterations in soluble guanylate cyclase content and modulation by nitric oxide in liver disease. *Neurochem Int* 2004;45:947–953.
 15. Feher J, Lengyel G, Blazovics A. Oxidative stress in the liver and biliary tract diseases. *Scand J Gastroenterol Suppl* 1998;228:38–46
 16. Furukawa Y. Histological changes in the brain due to experimental obstructive jaundice. *Nippon Geka Gakkai Zasshi* 1991;92:37–45.
 17. Gundersen HJ, Bagger P, Bendtsen TF, Evans SM, Korbo L, Marcussen N, et al. The new stereological tools Disector, fractionator, nucleator and point sampled intercepts and their use in pathological research and diagnosis. *APMIS* 1988;96:857-881.
 18. Huang LT, Chen CC, Sheen JM, Chen YJ, Hsieh CS, Tain YL. The interaction between high ammonia diet and bile duct ligation in developing rats: assessment by spatial memory and asymmetric dimethylarginine. *Int J Dev Neurosci* 2010;28:169-74.
 19. Huang LT, Tiao MM, Tain YL, Chen CC, Hsieh CS. Melatonin Ameliorates Bile Duct Ligation-Induced Systemic Oxidative Stress and Spatial Memory Deficits in Developing Rats. *Pediatr Res*. 2009;65:176-80.
 20. Huang YT, Hsu YC, Chen CJ, Liu CT, Wei YH. Oxidative-stress-related changes in the livers of bile-duct-ligated rats. *J Biomed Sci* 2003;10:170-178.
 21. Jover R, Rodrigo R, Felipe V, Insausti R, Saez-Valero J, Garcia-Ayllon MS, Suarez I, Candela A, Compan A, Esteban A, Cauli O, Auso E, Rodriguez E, Gutierrez A, Girona E, Erceg S, Berbel P, Perez-Mateo M. Brain edema and inflammatory activation in bile duct ligated rats with diet-induced hyperammonemia: A model of hepatic encephalopathy in cirrhosis. *Hepatology* 2006;43:1257–1266.
 22. Karaman A, Demirbilek S, Sezgin N, Gurbuz N, Gurses I. Protective effect of polyunsaturated phosphatidylcholine on liver damage induced by biliary obstruction in rats. *J Pediatr Surg* 2003;38:1341.
 23. Keller JN, Guo Q, Holsberg FW, Bruce-Keller AJ, Mattson MP. Increased sensitivity to mitochondrial toxin-induced apoptosis in neural cells expressing mutant presenilin-1 is linked to perturbed calcium homeostasis and enhanced oxyradical production. *J Neurosci* 1998;18:4439–4450.
 24. Keller JN, Mattson MP. Roles of lipid peroxidation in modulation of cellular signaling pathways, cell dysfunction, and death in the nervous system. *Rev Neurosci* 1998;9:105–116.
 25. Kirimlioglu H, Ecevit A, Yilmaz S, Kirimlioglu V, Karabulut AB. Effect of resveratrol and melatonin on oxidative stress enzymes, regeneration, and hepatocyte ultrastructure in rats subjected to 70% partial hepatectomy. *Transplant Proc* 2008;40:285-289.
 26. Lauderback CM, Hackett JM, Huang FF, Keller JN, Szweda LI, Markesbery WR, Butterfield DA. The glial glutamate transporter, GLT-1, is oxidatively modified by 4-hydroxy-2-nonenal in the Alzheimer's disease brain: the role of Abeta1–42. *J Neurochem* 2001;78:413–416.

27. Liu L, Liu H, Nam SW, Lee SS. Protective effects of erythropoietin on cirrhotic cardiomyopathy in rats. *Dig Liver Dis* 2012;44:1012-7.
28. Ljubuncic P, Tanne Z, Bomzon A. Evidence of a systemic phenomenon for oxidative stress in cholestatic liver disease. *Gut* 2000;47:710-716.
29. Ljubuncic P, Tanne Z, Bomzon A. Evidence of a systemic phenomenon for oxidative stress in cholestatic liver disease. *Gut* 2000;47:710-716.
30. Lockwood AH. Blood ammonia and hepatic encephalopathy. *Metab Brain Dis* 2004;19:345-349.
31. Low PA, Nickander KK, Tritschler H. The roles of oxidative stress and of antioxidant treatment in experimental diabetic polyneuropathy. *Diabetes* 1997;46:38-42.
32. Marchioli R. Antioxidant vitamins and prevention of cardiovascular disease: laboratory, epidemiological and clinical trial data. *Pharmacol Res* 1999;40:227-238.
33. Michiels C, Raes M, Toussaint O, Remacle J. Importance of Se-glutathione peroxidase, catalase, and Cu/Zn-SOD for cell survival against oxidative stress. *Free Radic Biol Med* 1994;17:235-248.
34. Mihara M, Uchiyama M. Effects of antioxidants on the TBA reaction of various rat liver homogenates, *Biochem Med* 1983;30:131-138.
35. Minor T, Isselhard W, Yamamoto Y, Obara M, Saad S. The effects of allopurinol and SOD on lipid peroxidation and energy metabolism in the liver after ischemia in an aerobic/anaerobic persufflation. *Surg Today* 1993;23:728.
36. Miyoshi H, Rust C, Roberts PJ, Burgart LJ, Gores GJ. Hepatocyte apoptosis after bile duct ligation in the mouse involves Fas. *Gastroenterology* 1999;117:669-677.
37. Moini H, Packer L, Saris NE. Antioxidant and prooxidant activities of alpha-lipoic acid and dihydrolipoic acid. *Toxicol Appl Pharmacol* 2002;182:84-90.
38. Morcos M, Borcea V, Isermann B et al. Effect of the antioxidant lipoic acid on the progression of endothelial cell damage and albuminuria in patient with diabetes mellitus. *Diabetes Res Clin Pract* 2001;52:175-183.
39. Mythili Y, Sudharsan PT, Amudha G, Varalakshmi P. Effect of DL-alpha-lipoic acid on cyclophosphamide induced lysosomal changes in oxidative cardiotoxicity. *Life Sci* 2007;80:1993-8.
40. Nagamatsu M, Nickander KK, Schmelzer JD et al. Lipoic acid improves nerve blood flow, reduces oxidative stress and improves distal nerve conduction in experimental diabetic neuropathy. *Diabetes Care* 1995;18:1160-1167.
41. Nickander KK, McPhee BR, Low PA, Tritschler H. Alpha-lipoic acid: antioxidant potency against lipid peroxidation of neural tissues in vitro and implications for diabetic neuropathy. *Free Radic Biol Med* 1996;21:631-639.
42. Onem G, Aral E, Y Enli, Oguz EO, Coskun E, Aybek H, et al. Neuroprotective Effects of L-Carnitine and Vitamin E Alone or in Combination Against Ischemia-Reperfusion Injury in Rats. *Journal of Surgical Research* 2006;131:124-130.
43. Orellana M, Rodrigo R, Thielemann L, Guajardo V. Bile duct ligation and oxidative stress in the rat: effects in liver and kidney. *Comp Biochem Physiol Toxicol Pharmacol* 2000;126:105-109.
44. Packer L, Tritschler HJ, Wessel K. Neuroprotection by the metabolic antioxidant α -lipoic acid. *Free Radical Biology and Medicine* 1996;22:359-378.
45. Packer L. α -Lipoic acid: a metabolic antioxidant which regulates NF- κ B signal transduction and protects against oxidative injury. *Drug Metabolism Reviews* 1998;30:245-275.
46. Paglia DE, Valentine WN. Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. *J Lab Clin Med* 1967;70:158-69.
47. Paoletti F, Aldinucci D, Mocali A, Caparrini A. A sensitive spectrophotometric method for the determination of superoxide dismutase activity in tissue extracts. *Anal Biochem* 1986;154:536-41.
48. Patel T, Bronk SF, Gores GJ. Increases of intracellular magnesium promote glycodeoxycholate-induced apoptosis in rat hepatocytes. *J Clin Invest* 1994;94: 2183-2192.
49. Perry G, Nunomura A, Hirai K, Zhu X, Perez M, Avila J, Castellani RJ, Atwood CS, Aliev G, Sayre LM, Takeda A, Smith MA. Is oxidative damage the fundamental pathogenic mechanism of Alzheimer's and other neurodegenerative diseases? *Free Radic Biol Med* 2002;33:1475-1479.
50. Portincasa P, Grattagliano I, Testini M, Caruso ML, Wang QH, Moschetta A, Calamita G, Vacca M, Valentini AM, Renna G, Lissidini G, Palasciano G. Parallel intestinal and liver injury during early cholestasis in the rat: modulation by bile salts and antioxidants. *Free Radic Biol Med* 2007;42:1381-139.
51. Quero JC, Schalm SW. Subclinical hepatic encephalopathy. *Semin Liver Dis* 1996;16:321-328.
52. Reed LJ, Debusk BG, Gunsalus IC, Hornberger CS. Crystalline α -lipoic acid: a catalytic agent associated with pyruvate dehydrogenase. *Science* 1951;114:93-94.

53. Rivera-Mancía S, Montes S, Méndez-Armenta M, Muriel P, Ríos C. Morphological changes of rat astrocytes induced by liver damage but not by manganese chloride exposure. *Metab Brain Dis* 2009;24:243-55.
54. Sastre J, Serviddio G, Pereda J, Minana JB, Arduini A, Vendemiale G, Poli G, Pallardo FV, Vina J. Mitochondrial function in liver disease. *Front Biosci* 2007;12:1200–1209.
55. Sayre LM, Smith MA, Perry G. Chemistry and biochemistry of oxidative stress in neurodegenerative disease. *Curr Med Chem* 2001;8:721–738.
56. Seyan AS, Hughes RD, Shawcross DL. Changing face of hepatic encephalopathy: role of inflammation and oxidative stress, *World Journal of Gastroenterology* 2010;3347–3357.
57. Shaafi S, Afrooz MR, Hajipour B, Dadadshi A, Hosseinian MM, Khodadadi A. Anti-oxidative effect of lipoic Acid in spinal cord ischemia/reperfusion. *Med Princ Pract* 2011;20:19-22.
58. Tokaç M, Taner G, Aydın S, Ozkardeş AB, Dündar HZ, Taşlıpınar MY, Arıkök AT, Kılıç M, Başaran AA, Basaran N. Protective effects of curcumin against oxidative stress parameters and DNA damage in the livers and kidneys of rats with biliary obstruction. *Food Chem Toxicol* 2013;S0278-6915(13):53-7.
59. Tomioka M, Iinuma H, Okinaga K. Impaired Kupffer cell function and effect of immunotherapy in obstructive jaundice. *J Surg Res* 2000;92:276.
60. Trauner M, Meierer PJ, Boyer JL. Molecular pathogenesis of cholestasis. *New Eng J Med* 1998;39:1217-1227.
61. Vasdev S, Stuckless J, Richardson V. Role of the immune system in hypertension: modulation by dietary antioxidants. *Int J Angiol* 2011;20:189-212.
62. Xie R, Li X, Ling Y, Shen C, Wu X, Xu W, Gao X. Alpha-lipoic acid pre- and post-treatments provide protection against in vitro ischemia-reperfusion injury in cerebral endothelial cells via Akt/mTOR signaling. *Brain Res* 2012;1482:81-90.
63. Ying Z, Kherada N, Farrar B, Kampfrath T, Chung Y, Simonetti O, Deiluiis J, Desikan R, Khan B, Villamena F, Sun Q, Parthasarathy S, Rajagopalan S. Lipoic acid effects on established atherosclerosis. *Life Sci* 2010;86:95-102.
64. Yuan GJ, Ma JC, Gong ZJ, Sun XM, Zheng SH, Li X. Modulation of liver oxidant-antioxidant system by ischemic preconditioning during ischemia/reperfusion injury in rats. *World J Gastroenterol* 2005;11:1825-1828.

2/27/2013