

## In Vitro Activity of nano-silver against Pulmonary Pathogenic Fungi

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**Abstract:** The in vitro activity of nano-silver versus those of amphotericin B was assessed against 37 plmonary aspergillois isolates. The activity of nano-silver against *Aspergillus* spp. is 2 times greater than that of amphotericin B. Nano-silver's antifungal activity was superior to hose of amphotericin B against plmonary pathogenic fungi in vitro.

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**Keywords:** nano-silver; Pulmonary aspergillois; drug susceptibility testing; antifungal

### 1. Introduction

Pulmonary aspergillois is a severe disease. Heretofore considered to be an unusual cause of infection, *Aspergillus* species have emerged as important causes of morbidity and mortality in immunocompromised patients (1-4). Invasive aspergillois currently constitutes the most common cause of infectious pneumonic mortality in patients undergoing HSCT(hematopoietic stem cell transplantation) and is an important cause of opportunistic respiratory and disseminated infection in other immunocompromised patients(4-11). Many risk factors are associated with Invasive aspergillois, a serious fungal infection that affects immunocompromised patients, particularly those with hematological malignancies and those who have undergone hematopoietic stem cell or solid organ transplantation.(12) Invasive fungus infections caused by *aspergillus* spp. occur most frequently in immunocompromised patients. A high infection-associated death rate of up to and over 50% is attributed even today to these fungi. The disease in humans is caused mainly by *Aspergillus fumigatus*, *Aspergillus flavus* and *Aspergillus niger*. Other species, for example, *Aspergillus terreus* or *Aspergillus nidulans* are quantitatively less prevalent. (13,14). Amphotericin B, a polyene macrolide, has long been used as the first-line agent for systemic fungal infection because of its broad spectrum. (15)

The silver ion is well known for its broad spectrum Antimicrobial at very low concentrations(16), And it has been used for centuries in health care delivery due to its antimicrobial and wound healing (anti-inflammatory) properties (17,18). Recently there is an increasing use of silver as an efficacious antibacterial and antifungal agent in wound care products and medical devices (10-22), and the advances in nanotechnology have enabled us to produce pure silver, as nanoparticles, which are more

efficient than silver ions (23). Nano-silver is available as an antimicrobial gel formulation for conventional topical antimicrobial agents, treatment (24). Some studies show that nano-silver has the antimicrobial activity against bacteria and virus (25-27). Our experiments have demonstrated that Nano silver exhibited potent antifungal activity against *Aspergillus* in vitro (28,29). The *Aspergillus* in our experiments such as *Aspergillus flavus*, *Aspergillus fumigatus*, *Aspergillus niger* are also the most common pathogens of Pulmonary aspergillois(13). In this study we want to determine the Activity of nano-silver against Pulmonary Pathogenic Fungi.

### 2. Material and Methods

Thirty-seven strains of *Aspergillus* isolated were obtained from patients with Pulmonary aspergillois from the Zhengzhou Central Hospital affiliated to Zhengzhou University, China, were investigated. These isolates were identified based on morphology by standard methods (30-33). Three species were studied, they included 16 *Aspergillus fumigatus*, 10 *Aspergillus flavus*, and 8 *Aspergillus niger*. *Candida parapsilosis* ATCC 22019 was used as quality control for each test. The antifungal agents tested in this study were nano-silver (Nanux, korea; 2000ppm) and amphotericin B (Bristol-Myers Squibb, Princeton, NJ) They were all dissolved in 100% dimethyl sulfoxide. The stock solutions were prepared at concentrations of 800µg/ml for nano-silver, 1,600 µg/ml for amphotericin B .Drug dilutions were made in RPMI 1640 (with L-glutamine, without sodium bicarbonate; GIBCO-BRL, Grand Island, NY) medium buffered to pH 7.0 with 0.165 M morpholinepropanesulfonic acid (MOPS; Serva, Feinbochemica GmbH, Germany). Final concentrations ranged from 0.0313 to 16µg/ml for nano-silver, from 0.0625 to 32µg/ml for amphotericin B. Then they were stored at -65°C until tested. A broth microdilution method was performed following the Clinical and Laboratory Standards

Institute (CLSI) M38-A document (34), which describes a standard method for testing the susceptibility of conidium-forming filamentous fungi that cause invasive fungal infections, including *Aspergillus* species, *Fusarium* species, etc., to antifungal agents. Inocula were prepared in accordance with the CLSI M38-A document. The final inoculum was  $0.4 \times 10^4$  to  $5 \times 10^4$  CFU/ml.

Following incubation at 35°C for 48 h, the MIC was determined according to the CLSI M38-A document. For both agents tested, the MIC was defined as the lowest drug concentration that prevented any discernible growth.

The MIC range and mode, the MIC50 (MIC for 50% of the strains tested), and the MIC90 (MIC for 90% of the strains tested) were provided for the isolates with the SPSS statistical package (version 13.0). For calculation, any high off-scale MIC was converted to the next higher concentration.

### 3. Results

The in vitro activities of nano-silver and

amphotericin B against the *Aspergillus* spp. are summarized in Table 1. The MIC50 and MIC90 of nano-silver were 0.5 µg/ml, 0.5 µg/ml, 0.25 µg/ml, respectively, and were 0.5 µg/ml, 1 µg/ml, 0.5 µg/ml, respectively, for *Aspergillus fumigatus*, *Aspergillus flavus* and *Aspergillus niger*. The MIC50 and MIC90 of amphotericin B were 1 µg/ml, 2 µg/ml, 1 µg/ml, respectively, and were all 2 µg/ml for *Aspergillus fumigatus*, *Aspergillus flavus* and *Aspergillus niger*. When comparing the MIC90s of nano-silver and amphotericin B, the activity of nano-silver against *Aspergillus* spp. is 2 times greater than that of amphotericin B. And as shown in Tables 1, nano-silver has activity against *Aspergillus* complexes. For each of these genera, this activity remains consistent and does not show significant interspecies variability. Therefore, nano-silver was effective against main Pulmonary pathogenic fungi in vitro. And its effect was superior to those of amphotericin B.

Tables 1. In vitro susceptibilities of Pulmonary *Aspergillus* isolates to Nano-silver and amphotericin B.

Organism (no. of isolates) and antifungal agent	MIC range (µg/ml)	MIC mode (µg/ml)	MIC50 (µg/ml)	MIC90 (µg/ml)
<i>Aspergillus fumigatus</i> species complex (16)				
Nano-silver	0.25-1	0.5	0.5	0.5
amphotericin B	0.5-4	1	1	2
<i>Aspergillus flavus</i> species complex (10)				
Nano-silver	0.5-1	0.5	0.5	1
amphotericin B	1-32	2	2	2
<i>Aspergillus niger</i> species complex (8)				
Nano-silver	0.125-0.5	0.5	0.25	0.5
amphotericin B	0.25-2	1	1	2
<i>Aspergillus</i> spp.(34)				
Nano-silver	0.125-1	0.5	0.5	1
amphotericin B	0.25-32	1	1	2

### 4. Discussions

The scientific literature points that Nano-silver is widely used in medical devices and supplies as a potent antibacterial, antifungal, antiviral, and anti-inflammatory agent. (35,36). Coatings generally comprised of nanoparticles have been used to prevent bacterial infections associated with medical devices, such as wound dressings, catheters, and orthopedic and cardiovascular implants, with different degrees of clinical efficacy (37,38). The findings from our study indicate that nano-silver is active against main Pulmonary pathogenic fungi. The results suggest that a prospective evaluation of efficacy and safety to develop the nano-silver's clinical applications such as Fiberoptic bronchoscopy.

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### References

- Patterson TF, Kirkpatrick WR, White M, et al. Invasive aspergillosis: disease spectrum, treatment practices, and outcomes. I3 *Aspergillus* Study Group. *Medicine* (Baltimore) 2000; 79:250-260.
- Denning DW. Invasive aspergillosis. *Clin Infect Dis* 1998; 26:781-803.
- Marr KA, Patterson T, Denning D. *Aspergillosis: pathogenesis, clinical manifestations, and therapy.* *Infect Dis Clin North Am* 2002; 16:875-894.

4. Walsh TJ, Anaissie EJ, Denning DW, Herbrecht R, Kontoyiannis DP, Marr KA, Morrison VA, Segal BH, Steinbach WJ, Stevens DA, van Burik JA, Wingard JR, Patterson TF. Treatment of aspergillosis: clinical practice guidelines of the Infectious Diseases Society of America. *Clin Infect Dis*. 2008 Feb 1; 46(3):327-360.
5. Cornet M, Fleury L, Maslo C, Bernard JF, Brucker G. Epidemiology of invasive aspergillosis in France: a six-year multicentric survey in the greater Paris area. *J Hosp Infect* 2002;51:288-296
6. Benjamin DK Jr, Miller WC, Bayliff S, Martel L, Alexander KA, Martin PL. Infections diagnosed in the first year after pediatric stem cell transplantation. *Pediatr Infect Dis J* 2002; 21:227-234.
7. Grow WB, Moreb JS, Roque D, et al. Late onset of invasive aspergillus infection in bone marrow transplant patients at a university hospital. *Bone Marrow Transplant* 2002;29:15-19
8. Marr KA, Carter RA, Boeckh M, Martin P, Corey L. Invasive aspergillosis in allogeneic stem cell transplant recipients: changes in epidemiology and risk factors. *Blood* 2002; 100:4358-4366.
9. Montoya JG, Chaparro SV, Celis D, et al. Invasive aspergillosis in the setting of cardiac transplantation. *Clin Infect Dis* 2003;37 (Suppl 3):281-292
10. Paterson DL, Singh N. Invasive aspergillosis in transplant recipients. *Medicine (Baltimore)* 1999; 78:123-138.
11. Wald A, Leisenring W, van Burik J-A, Bowden RA. Epidemiology of Aspergillus infections in a large cohort of patients undergoing bone marrow transplantation. *J Infect Dis* 1997; 175:1459-1466.
12. Denning DW. Invasive aspergillosis. *Clin Infect Dis* 1998; 26:781-803.
13. Karthaus M. Guideline based treatment of invasive aspergillosis. *Mycoses*. 2010 May; 53 Suppl 1:36-43.
14. Jeannina A. Smith And Carol A. Kauffman. Pulmonary fungal infections. *Respirology*. Volume 17, Issue 6, August 2012.913-926
15. Nagasaki Y, Eriguchi Y, Uchida Y, Miyake N, Maehara Y, Kadowaki M, Harada M, Akashi K, Shimono N. Combination therapy with micafungin and amphotericin B for invasive pulmonary aspergillosis in an immunocompromised mouse model. *J Antimicrob Chemother*. 2009 Aug; 64(2):379-382.
16. Melaiye A, Youngs WJ. Silver and its application as an antimicrobial agent. *Expert Opin*. 2005; 15:125-130.
17. Chaloupka K, Malam Y, Seifalian AM. Nanosilver as a new generation of nanoproduct in biomedical applications. *Trends Biotechnol*. 2010; 28:580-588.
18. Lansdown ABG. A pharmacological and toxicological profile of silver as an antimicrobial agent in medical devices. *Adv. Pharmacol. Sci*. 2010; 2010:1-16.
19. Kim JS, Kuk E, Yu KN, Kim JH, Park SJ, Lee HJ, Kim SH, Park YK, Park YH, Hwang CY, Kim YK, Lee YS, Jeong DH, Cho MH. Antimicrobial effects of silver nanoparticles. *Nanomedicine*. 2007; 3:95-101.
20. Lara HH, Ayala-Nuñez NV, Ixtapan-Turrent L, Rodriguez-Padilla C. Bactericidal effect of silver nanoparticles against multidrug-resistant bacteria. *World Journal of Microbiology and Biotechnology*. 2010; 26:615-621.
21. Salata O. Applications of nanoparticles in biology and medicine. *J Nanobiotechnology*. 2004 Apr 30; 2(1):3.
22. Shahverdi AR, Fakhimi A, Shahverdi HR, Minaian S. Synthesis and effect of silver nanoparticles on the antibacterial activity of different antibiotics against *Staphylococcus aureus* and *Escherichia coli*. *Nanomedicine*. 2007; 3:168-171.
23. Lara HH, Ayala-Nunez NV, Ixtapan-Turrent L, Rodriguez-Padilla C. Mode of antiviral action of silver nanoparticles against HIV-1. *J Nanobiotechnology*. 2010 Jan 20; 8:1.
24. Muangman P, Chuntrasakul C, Silthram S, Suvanchote S, Benjathanung R, Kittidacha S, Rueksomtawin S. Comparison of efficacy of 1% silver sulfadiazine and Acticoat for treatment of partial-thickness burn wounds. *J Med Assoc Thai*. 2006; 89(7):953-958.
25. Chen M, Yang Z, Wu H, Pan X, Xie X, Wu C. Antimicrobial activity and the mechanism of silver nanoparticle thermosensitive gel. *Int J Nanomedicine*. 2011; 6: 2873-2877.
26. Galdiero S, Falanga A, Vitiello M, Cantisani M, Marra V, Galdiero M. Silver nanoparticles as potential antiviral agents. 2011 Oct 24; 16(10): 8894- 8918.
27. Nanda A, Saravanan M. Biosynthesis of silver nanoparticles from *Staphylococcus aureus* and its antimicrobial activity against MRSA and MRSE. *Nanomedicine*. 2009 Dec; 5(4):452-456.
28. Yan Xu, Guangren Pang, Chuanwen Gao, Dongqing Zhao, Lutan Zhou, Shengtao Sun, and Bingliang Wang. In Vitro Comparison of the Efficacies of Natamycin and Silver Nitrate against Ocular Fungi. *Antimicrob Agents Chemother*. 2009 Apr; 53(4):1636-1638.
29. Chuanwen Gao, Yan Xu, Chao Xu. In Vitro Activity of nano-silver against Ocular Pathogenic Fungi. *Life Science Journal*. 2012; 9(4):750-753.

30. Wang LY, Sun ST, Zhu L, Zhang YQ, Wang YQ, Li JC, and Xu J. The pathogenic spectrum investigation of fungal keratitis in 1996\_2002 of Henan. *Chin. J. Pract. Ophthalmol.* 2003; 21:224-225.
31. Wang LY, Zhang YQ, Wang YQ, Wang GS, Lu JB, and Deng JH. Spectrum of mycotic keratitis in China. *Chin. J. Ophthalmol.* 2000; 36:138-140.
32. Wei JC. Identification manual of fungi. Scientific & Technologic Press, Shanghai, China. 1977. *Aspergillus Micheli ex Fr.*, p. 495–500. In J. C. Wei (ed.)
33. Sun ST, Wang LY, Wang GS, Zhou Y, Zhang YQ, Zhu L, and Deng JH. Spectrum of 90 cases with mycotic keratitis. *Chin. Ophthalmic Res.* 2002; 20:247-248.
34. National Committee for Clinical Laboratory Standards. 2002. Reference method for broth dilution antifungal susceptibility testing of filamentous fungi. Approved standard M38-A. National Committee for Clinical Laboratory Standards, Wayne, PA.
35. Carmen Steluta Ciobanu, Florian Massuyeau, Liliana Violeta Constantin, and Daniela Predoi Structural and physical properties of antibacterial Ag-doped nano-hydroxyapatite synthesized at 100°C *Nanoscale Res Lett.* 2011; 6(1): 613.
36. Pankhurst QA, Connolly J, Jones SK, Dobson J. Applications of magnetic nanoparticles in biomedicine. *J Phys D: Appl Phys.* 2003; 36:R167.
37. Wijnhoven SWP, Peijnenburg WJGM, Herberts CA, Hagens WI, Oomen AG, Heugens EHW, Roszek B, Bisschops J, Gosens I, Van De Meent D. Nano-silver—a review of available data and knowledge gaps in human and environmental risk assessment. *Nanotoxicology* 2009;3:109-138.
38. Choi J, Reipa V, Hitchins VM, Goering PL, Malinauskas RA. Physicochemical characterization and in vitro hemolysis evaluation of silver nanoparticles. *Toxicol Sci.* 2011 Sep; 123(1):133-143.

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