New Aspect in Cartilage Growth "The Invasive Interstitial Type"

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Abstract: Growth of hyaline cartilage is achieved either by chondrocyte division "interstitial type of growth" or by perichondrial cell differentiation of mesenchymal cells to differentiated chondrocytes which deposit successive circumferential layers of cartilage matrix "appositional growth". The current study was carried out to illustrate a third type of cartilage growth during development of the air breathing dendritic organ of catfish. To perform this study, samples of the air breathing dendritic organ were collected from catfish (Clarias garie-pinus). Paraffin sections of formalin fixed specimens were examined by light microscopy. During development of the air breathing organ of catfish, the nidus of cartilage formation was represented by a condensation of mesenchymal cells. Some cells differentiate to chondrocytes to form multiple cartilage islets surrounded by mesenchymal tissue which was enclosed by perichondrium. The surrounding mesenchymal cells further on differentiate to chondrocytes to complete the developing elastic cartilaginous unit. Additional mesenchymal cells with chondrocyte potential invade the cartilage at multiple sites to grow the cartilaginous mass interstitially. Moreover, interstitial mesenchymal differentiation occurred at the sites of the vacated lacunae which resulted from the death of chondrocytes. Interstitial mesenchymal invasion occurred in the hyaline cartilage to be replaced by elastic type. The invasive interstitial differentiation of the mesenchymal cells contributed in cartilage growth, renewal, and replacement. Studying the molecular regulation of the invasive differentiation of the mesenchymal cells may be a useful guide in researches to resolve cartilage disorders problems.

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1. Introduction

Embryonic mesenchymal progenitors are multipotent cells and exhibit the potential to differentiate into multiple cell lineages. Mesenchymal cells destination depends on the molecular environment that enable cells to acquire phenotypic characteristics and perform the tissue functional requirements. Mesenchymal cells can differentiate to chondrogenic or osteogenic cells to form the skeletal elements. During development of cartilaginous elements, the undifferentiated mesenchymal cells transform to chondrocytes. Chondrocytes secrete specific cartilage proteins and form a cartilage template. Cartilage growth depends on the activities of chondrocytes and preichondrial stem cells. Interstitial division of chondrocytes commits elongation of the cartilage tissue "interstitial growth of the cartilage". Perichondral cells differentiate to chondrocytes and lay down circumferential layers of cartilage matrix to increase the cartilage diameter "appositional growth" (Singh, 2010). Embryonic mesenchymal cells invasion to cartilage template and further differentiation has been reported in a previous study (Soliman, 2013). A similar mode of cartilage growth was described in the present investigation. The current study was performed in the elastic cartilage of the air breathing organ in catfish to clarify the difference between growth of mammalian cartilage and the interstitial mesenchymal differentiation during cartilage growth in the air breathing dendritic organ of catfish.

2. Material and methods

The study was carried out on 8 apparent healthy catfish of different ages. Juvenile cat fishes (*Clarias garie-pinus*) were caught from the River Nile and transported in a glass pool in the lab. Samples of the air breathing organ (suprabranchial organ) were collected from 13- 27 cm measured catfish (Table 1). The obtained samples were fixed in 10% formalin buffered solution, dehydrated and impregnated and embedded in paraffin wax. Longitudinal sections of 3-5 μ m were undertaken and mounted on glass slides. The sections were kept in incubator at 40°C, stained with H&E and Safranin O and examined by light microscopy.

Table 1	: length	of the	developing	catfish
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Early stages of developing catfish (cm)	Advanced stages of developing catfish (cm)	
13.4	20.9	
13.9	24.0	
15.4	26.3	
16.2	27.2	

3. Results

The air respiratory organ of the catfish appeared as arborizing-like structure associated with the gills and located in the suprabranchial air chamber (Fig. 1). Development of this organ depends to great extent on formation of the supporting cartilage units. In the arborizing terminals, the primordia of cartilage elements were formed. The first sign of initiation of cartilage elements is condensation of the mesenchymal cells (Figs 2A, B). Some mesenchymal cells differentiate to chondrocytes. The primitive suprabranchial cartilage appears as islets of cartilaginous mass embedded in mesenchymal tissue surrounded by perichondrium. Mesenchymal cells later on transformed to chondrocytes (Figs 2C, D, E) and deposit a cartilage matrix to form a large cartilage mass. Undifferentiated mesenchymal cells originated from the perichondrium, penetrated the interior of the cartilaginous mass and later on transformed to chondrocytes (Fig 2G). Vascular cells originating from the vascular rich subepithelial connective tissue penetrate the perichondrium and invade the cartilage in association with mesenchymal cells (Fig 2F). The cellular invasion was confined to certain areas in the cartilage and was responsible for the unique enlargement of the cartilaginous mass giving the irregularity of the cartilaginous surface (Fig 3A).

The perichondral cells differentiated to chondrocytes to grow the cartilage appositionally (Fig 3F). Moreover, the invasive undifferentiated mesenchymal cells nearby the perichondrium differentiated to chondrocytes (Fig 3E).

With advancement of age, some hypertrophied chondrocytes reached their terminal differentiation and die leaving vacated lacunae (Figs 3B,D). Mesenchymal cells invaded the growing cartilage to occupy the vacated areas and differentiate to chondrocytes (Fig 3B). During cellular invasion, mesenchymal cells can be observed breaking down the septal matrix and opened the lacunae (Figs 5A-F). Various degrees of differentiation and hypertrophy of chondrocytes and vacated lacunae can be observed in the cartilage (Fig 3D). Chondrocytes doublets were rarely seen (Fig 2E).

In certain locations, cartilage was invaded by vascular cells and the surrounding dense perichondrial tissue to divide it into cartilaginous units (Figs 4A, C) where the covering epithelium invaginated (Figs 4B, D).

At the base of air breathing organ, hyaline cartilage was formed and gradually was replaced by elastic type by the mode of invasive interstitial growth. Mesenchymal cells invade the hyaline cartilage and differentiate to chondrocytes capable of synthesizing and secreting elastic fibers (Figs 6 A, B, C, D, E).

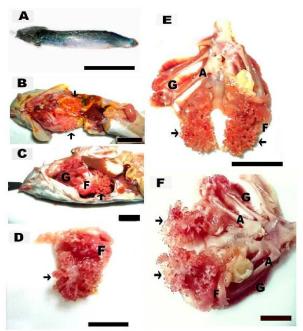


Figure 1: Gross anatomy of the air respiratory organ in catfish

A: photograph of catfish (Clarias garie-pinus). B: Ventral view of the catfish illustrated the air breathing dendritic organ. C: lateral view of the catfish shows gills and the associated air breathing organ D: shows the air breathing dendritic organ. E,F: respiratory system of catfish. Note: The arrows refer to the air breathing organ. Gill fan (F), Gill filaments (G) and Gill arches (A). Scale bars represent 5cm in "A", 2cm in "B, C", 1 cm in "D-E".

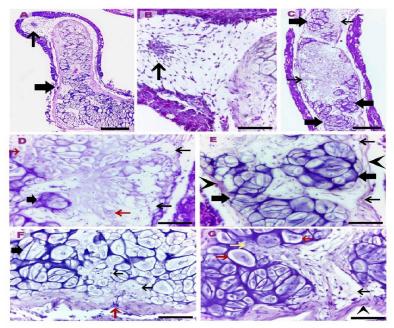
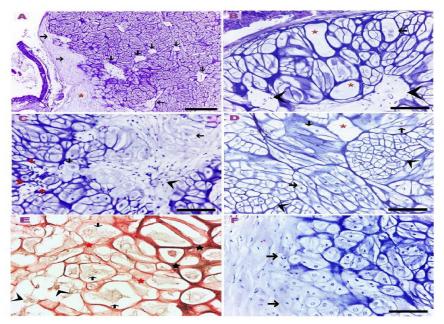
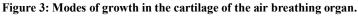


Figure 2: Early differentiation of the cartilaginous support of the air breathing organ

Paraffin sections of air respiratory organ of catfish measured 13.4 (A), 13.2 (C-E), 14.1 (F), 14.9 (G) cm in length, stained with H&E. A, B: The thin arrows refer to the areas of mesenchymal cell condensation. C: The thin arrow point to the differentiating chondrocytes; Note little extracellular cartilage matrix around the differentiating chondrocytes. The thick arrows refer to the cartilaginous masses. D, E, G: The thin arrows refer to areas of the cellular invasion to the growing cartilage. The red arrows refer to differentiating chondrocytes in "D", chondrocytes doublets in "G" and the invading perichondrial mesenchymal cells to the growing cartilage in "F". The yellow arrow refers to the differentiating chondrocytes. The arrow heads refer to the perichondrium. Scale bars represent 80 µm in "A, C", 50 µm in "B, D-G".





Paraffin sections of air respiratory organ of catfish measured 20.9 (A), 27.3 (B), 14.1 (C), 26.3 (D), 14.9 (G) cm in length, stained with H&E (A-D, F) and safranin O (E). The thin arrows point to the sites of cellular invasion in "A", terminally differentiated chondrocyte in "B", differentiating chondrocytes in "C, E", large groups of chondrocytes in "D" and area of appositional growth in "F". The red arrows refer to the areas of cartilage remodeling. The red asterisks located in the vacated lacunae "B, D" and refer to proteoglycan poor matrix "E". The black asterisks refer to proteoglycan rich matrix "E". The arrow heads refer to the invading mesenchymal cells in "B, C, E". Scale bars represent 80 µm in "A", 50 µm in "B-D, F", 20 µm in "E".

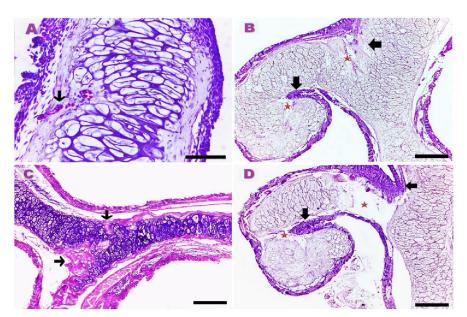


Figure 4: Divisions of cartilaginous units and formation of respiratory compartments in air breathing organ. Paraffin sections of air respiratory organ of catfish measured 26.3 (A), 24.6 (B, D), 20.9 (C)cm in length, stained with H&E. The thin arrows refer to areas of vascular and perichondrial CT invasion into the cartilage. The thick arrows refer to epithelial invaginations. The red asterisks refer to divisions of cartilaginous units. Scale bars represent 50 µm in "A", 80 µm in "B-D".

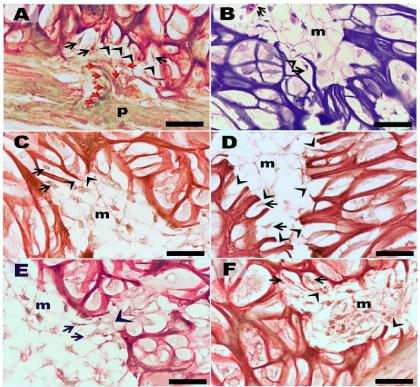


Figure 5: Degradation of cartilage matrix during mesenchymal invasion.

Paraffin sections of air respiratory organ of catfish measured 24.6 (A, B), 26.3 (C, F), 20.9 (D, E) cm in length, stained with safranin O "A, C- F" and H&E "B". The black arrows refer to penetration of mesenchymal cells into lacunae. The red arrows refer to the perichondral originating undifferentiated cells. The arrow heads refer to the opened lacunae. The double arrow refers to vascular invasion. m: mesenchymal cells. Scale bars represent 20 µm

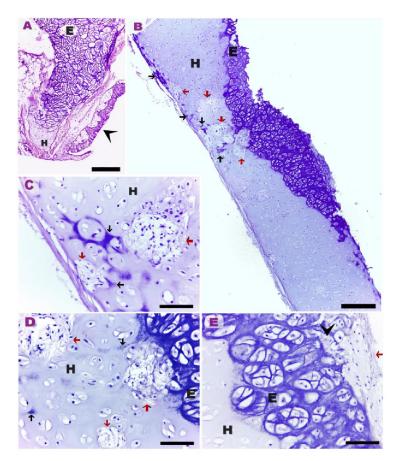


Figure 6: Replacement of hyaline cartilage by elastic type in air breathing organ.

Paraffin sections of air respiratory organ of catfish measured 24.6 (A), 26.3 (B-E) cm in length, stained with H&E. The arrow head refers to the respiratory epithelium in "A" and differentiating chondrocytes of elastic cartilage in "E". The red arrows refer to areas of interstitially located mesenchymal cells. The black arrows refer to the matrix of the newly formed elastic cartilage. Note: "H" hyaline and "E" elastic cartilage. Scale bars represent 80 µm in "A", 200 µm in "B", 20 µm in "C" 50 µm in "D"

4. Discussion

Developmental events of skeletal elements in mammals and avian species have been investigated in many previous studies; however Aquarian species had not taken a great interest. In the current investigation, I will focus on an unusual mode of cartilaginous development and growth in the dendritic air breathing organ in catfish and compared with the sequential events and commonly known modes of cartilage growth. In the present study, Mesenchymal cell condensation marked the initial site of cartilage development. Multiple focal areas of mesenchymal cells differentiate to chondrocytes to cartilaginous islets interspersed among form undifferentiated mesenchyme and surrounded by a delicate perichondrium. Unlike the mammalian and avian species, mesenchymal cell condensation was followed by cartilage primordia which contain prechondrocytes at the same stage of differentiation and activity secreting the cartilage matrix and surrounded by perichondrial cells (Singh, 2010 and Soliman, 2013).

Growth and chondrocytes orientation in cartilage in the air breathing organ was unusual. During cartilaginous growth, multiple sites of cellular invasion can be detected where undifferentiated mesenchymal cells transformed to chondrocytes and secrete cartilage matrix. However, interstitial growth of chondrocytes in the cartilages of the dendritic respiratory organ was rarely observed and isogenous groups or cell nests were not the usual form of chondrocytic organization. Unlike mammals and avian species, interstitial cartilaginous growth of air breathing organ depended primarily on the interstitial invasion and differentiation of the mesenchymal cells. Perichondrial cells contributed in appositional growth in co-ordination with peripheral mesenchymal cells differentiation. Unlike mammals, appositional growth occurs only through the activities of the perichondrial cells (Martini, 2007 and Singh, 2008). differentiation The invasive interstitial of mesenchymal cells resulted in a characteristic organization of hypertrophied chondrocytes in the cartilage of the air breathing organ as they were observed in groups of small and large hypertrophic chondrocytes and vacated lacunae. A different organization is usually observed in mammalian cartilages. Appositional growth is responsible for arrangement of the immature chondrocytes peripherally and gradually chondrocytes undergo maturation toward the center of the cartilaginous core (Martini, 2007).

Invasive mesenchymal differentiation was recorded in a previous study during growth of cartilage templates of the femur and tibia in quail embryos. Mesenchymal cells of periosteal origin invade the interior of the cartilage template. The cellular invasion was confined to the central hypertrophic zone of the cartilage template and was not observed in the proliferating, resting zones or articular cartilage. Mesenchymal invasion aims to provide the cartilage templates with a population of chondrogenic cells to increase the cartilage diameter of the hypertrophic region (Soliman, 2013). It should mentioned that mesenchymal osteogenic be progenitors usually invade the hypertrophic zone during endochondral ossification (Carlevaro et al., 2000).

In the largest cartilaginous units of the dendritic air breathing organ. hypertrophic chondrocytes died and empty lacunae were invaded by mesenchymal cells which transformed to chondrocytes. The aim of the cellular invasion may be to regenerate and continue renewal of the cartilaginous tissue. The present study suggests that the mesenchymal cells can degrade the matrix in the invasive pathway. Degeneration of elastic cartilage of the air breathing organ represented by break down of the thin septal matrix, unlike the abundant matrix in hyaline cartilage loss the affinity of staining with proteoglycan reacting stain. The proteolytic activity of the mesenchymal cells has been studied by (Ghajar et al., 2010) and is required for division and growth of the tissues (Shi et al., 2008).

Certain areas of cellular invasion carried the vascular branches from the vascular rich subepithelial connective tissue to the interior of the cartilaginous tissue. The dense perichondrial connective tissue and the associated vascular cells and epithelial covering penetrate the cartilage divide it into masses. This type of invasion may aim to redistribute new vascular elements and the prospective respiratory lamellar epithelial covering to increase the respiratory surface and create new respiratory compartments to adopt the respiratory function.

In the present study, the interstitial differentiation of mesenchyaml cells was observed in the basal hyaline cartilage in the dendritic respiratory organ. Cellular invasion provides the hyaline cartilage with chondrogenic cells which can deposit elastic fibers. The elastic matrix has been detected by Verhoeff's elastic stain (Ahmed *et al.*, 2008).

In conclusion, the dynamics of mesnchymal tissue play a central role during development of the cartilaginous support of the dendritic air breathing organ. Invasive interstitial differentiation of the mesenchymal cells was responsible for growth, regeneration and replacement of the cartilage.

Clinical correlations

In many skeletal disorders, cartilage pathology is dominated by changes in the matrix. In articular cartilage poor regenerative capacity as a response to matrix injury and matrix loss is now well documented. The use of mesenchymal or stem cell technology to improve this situation is currently of great interest. This study demonstrates the capacity of mesenchymal cells with chondrogenic potential to existing cartilage matrix during replace an development. This study should give encouragement to those who are working with stem cell technologies that under special circumstances stem cell technology to repair articular hyaline cartilage may be a clinical possibility. Cartilage of the catfish respiratory organs can be used.

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