Determination of the dominant families in Ilkhji region, Eastern Azerbaijan province (Northwest of Iran)

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Abstract: Systematic Knowledge or plant taxonomy is one of the very old and important branches of botany. By gathering plants, we can recognize rare species of plants or those which are facing extinction, so that we can find some ways to prevent them from destruction. In this research, all plants were gathered from Ilkhji area which is located in East Azerbaijan province. Ilkhji region is located in 25 km south west of Tabriz and the geographic coordinates 45.59 to 12 and 46.3 eastern longitudes and 37.55 to 37.57 north latitude. Plant samples from Yal, Khaselar, Kordlar and Chaman areas as well, were obtained during winter of year 2008 to fall year 2009. Standard method was followed with regard to collection of plant materials, drying, mounting, preparation and preservation of plant specimens' .All the plant samples were pressed according to standard guides. If the plant samples were too long, then they were cut from several areas, so the sample contained the complete plant. At the next stage, samples were stick to the herbarium Cardboards and then were identified using floras, keys, illustrations and explanations which are available for different sources of plant Species. Dominant plant families consist of: Asteraceae (30 species), Brassicaceae (23 species), Fabaceae (21 species), Poaceae (21 species) Boraginaceae (16 species) and Lamiaceae (16 species).

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Key words: herbarium, systematic, flora.

Introduction

Plant systematic science provide tools to make a list of plants, the methods of identification, name and ordering, however, can be the basic knowledge for the biological science. Identification of a plant, introduces a specific way to determine the natural condition of that and bring about an introduction to recognize the relationship between different species. Collecting plants or endangered plants likely helps to improve methods to save them and also revealers their far relationships with the well-known species. In sum, dominant species greatly affect both physical and biological conditions, and it makes sense to use them to examine community condition. The behavior of a dominant species and its relationship to other species are not necessarily constant, however. A dominant species can be highly competitive in a core habitat or able to tolerate stress in a peripheral habitat (Wisheu and Keddy 1992). Substantial information about both abiotic and biotic properties of a plant community is conveyed simply by identifying the dominant species (as in Clements 1916, Whittaker 1965). Through their architecture, physiology, growth, and phonology, dominant plants determine overall community structure, such as biomass and canopy strata (Richards 1996) and ecosystem engineering (Malmer et al. 2003); soil properties (Bardgett et al. 1999); ways of succession (Fastie 1995); ecosystem properties, such as nutrient cycling (Allison and Vitousek 2004) and fire regimes (Taylor 2000); micro-habitats for subordinate species (Grime 1998); and even hydrological conditions (Simberloff and Von Holle 1999). Dominant plants can exert strong influence by their abundance, height, shade, root and rhizome biomass, or chemistry (e.g., allelopathy). Iran, has a diversity of plant varieties which because of the specific geographic locations, great number of them are not known, therefore, the identification and name these plants is of a great importance.

2. Materials and Methods

All the plant samples in this research were gathered from Ilkhji area which is located in East Azerbaijan province. Ilkhji area is located in 25 km south west of Tabriz and the geographic coordinates 45.59 to 12 and 46.3 Eastern longitudes and 37.55 to 37.57 North latitude. Plant samples from Yal, Khaselar, Kordlar and Chaman areas as well, were obtained during winter of year 2008 to fall in the year 2009. Standard method was followed with regard to collection of plant materials, drying, mounting, preparation and preservation of plant specimens (Shrestha and Dhillion, 2003). All the plant samples were pressed according to standard guides. If the plant samples were too long, then they were cut from several areas, so the sample contained the complete plant. At the next stage, samples were stick to the herbarium cardboards and they were identified using floras, keys, illustrations and explanations which are available for different sources of plant Species. Finally dominant plants were separated and introduced.

RESULTS

Result of survey show that Dominant plant families consist of: Asteraceae (30 species), Brassicaceae (23 species), Fabaceae (21 species), Poaceae (21 species) Boraginaceae (16 species) and Lamiaceae (16 species). Results showed as Tables 1.

| Family | Genus | Species | Growth habite |
|--------------|---------------|------------------|-----------------|
| Brassicaceae | Alyssum | A.bracteatum | Hemicriptophite |
| | Alyssum | A.dasycarpum | Hemicriptophite |
| | Alyssum | A.longistylum | Hemicriptophite |
| | Alyssum | A.linifolium | Hemicriptophite |
| | Alyssum | A.minus | Hemicriptophite |
| | Capsella | C.bursa-pastoris | Therophite |
| | Cardaria | C.draba | Hemicriptophite |
| | Choriospora | Ch.tenella | Therophite |
| | Conringia | C.perfoliata | Therophite |
| | Descurainia | D.sophia | Hemicriptophite |
| | Erysimum | E.aitchisonii | Hemicriptophite |
| | Erysimum | E.crassipes | Hemicriptophite |
| | Erysimum | E.cuspidatum | Hemicriptophite |
| | Erysimum | E.deifolium | Hemicriptophite |
| | Erysimum | E.filifolium | Hemicriptophite |
| | Lepidium | L.perfoliatum | Hemicriptophite |
| | Malcolmia | M.africana | Therophite |
| | Malcolmia | M.iberica | Therophite |
| | Raphanus | M.raphanistrum | Therophite |
| | Strigmostemum | S.sulphureum | Hemicriptophite |
| | Sisymberium | S.loeselii | Hemicriptophite |
| | Thlaspi | T.arvense | Therophite |

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| | Thlaspi | T.umbellatum | Therophite |
|---------------|--------------|-----------------|-----------------|
| Papilionaceae | Alhaji | A.camelo | Hemicriptophite |
| | Astragalus | A.askius | Hemicriptophite |
| | Astragalus | A.chrysostachys | Hemicriptophite |
| | Astragalus | A.effuses | Hemicriptophite |
| | Astragalus | A.holopsilus | Hemicriptophite |
| | Astragalus | A.oroboides | Therophite |
| | Astragalus | A.globiflorus | Therophite |
| | Coronilla | C.balansae | Therophite |
| | Coronilla | C.varial | Therophite |
| | Hedysarum | H.formosum | Therophite |
| | Lotus | L.corniculatus | Hemicriptophite |
| | Medicago | M.sativa | Hemicriptophite |
| | Melilotus | M.officinalis | Therophite |
| | Onobrychis | O.cornuta | Therophite |
| | Trifolium | T.canescens | Therophite |
| | Trifolium | T.clusii | Therophite |
| | Trifolium | T.repens | Therophite |
| | Trigonella | T.aurantiaca | Therophite |
| | Trigonella | T.coerulescens | Hemicriptophite |
| | Trigonella | T.monantha | Hemicriptophite |
| | Trigonella | T.monspeliaca | Hemicriptophite |
| Boraginaceae | Alkanna | A.orientalis | Hemicriptophite |
| | Alkanna | A.brateosa | Hemicriptophite |
| | Anchusa | A.italica | Hemicriptophite |
| | Asperugo | A.procumbens | Therophite |
| | Heliotropium | H.brevilimbe | Therophite |

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| | Heliotropium | H.swtanense | Hemicriptophite |
|-----------|--------------|------------------|-----------------|
| | Heterocaryum | H.szovitsianum | Therophite |
| | Lappula | L.sinaica | Therophite |
| | Lithospermum | L.arvense | Hemicriptophite |
| | Moltkia | M.coerulea | Camephite |
| | Moltkia | M.grpsacea | Hemicriptophite |
| | Nonnea | N.caspica | Camephite |
| | Nonnea | N.persica | Camephite |
| | Onosma | O.kotschyi Boiss | Camephite |
| | Rochelia | R.disperma | Camephite |
| | Rochelia | R.persica | Camephite |
| Lamiaceae | Marrubium | M.vulgare | Geophite |
| | Mentha | M.longifolia | Hemicriptophite |
| | Nepeta | N.meyeri | Hemicriptophite |
| | Nepeta | N.persica | Hemicriptophite |
| | Nepeta | N.racemosa | Hemicriptophite |
| | Phlomis | Ph.olivieri | Hemicriptophite |
| | Salvia | S.nemorasa | Hemicriptophite |
| | Salvia | S.sahendica | Hemicriptophite |
| | Salvia | S.spinosal | Hemicriptophite |
| | Salvia | S.virgata | Hemicriptophite |
| | Stachys | S.inflata | Hemicriptophite |
| | Stachys | S.lavandifolia | Hemicriptophite |
| | Stachys | S.turcomanica | Hemicriptophite |
| | Thymus | T.cotschyanus | Geophite |
| | Thymus | T.pubescens | Geophite |
| | Ziziphora | Z.tenuior | Therophite |

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| Asteraceae | Achillea | A.millefolium | Hemicriptophite |
|------------|--------------|----------------|-----------------|
| | Achillea | A.vermicularis | Hemicriptophite |
| | Acroptilon | A.repens | Hemicriptophite |
| | Anthemis | A.hyalina | Therophite |
| | Calendula | C.persica | Therophite |
| | Carthamus | C.oxyacantha | Therophite |
| | Centaurea | C.depressa | Hemicriptophite |
| | Centaurea | C.pulchella | Hemicriptophite |
| | Centaurea | C.virgata | Hemicriptophite |
| | Cichorium | C.intybus | Hemicriptophite |
| | Cirsium | C.arvense | Hemicriptophite |
| | Cirsium | C.congestum | Hemicriptophite |
| | Cnicus | C.benedictus | Therophite |
| | Cosinia | C.rhaphiostega | Camephite |
| | Crepis | C.foetida | Hemicriptophite |
| | Crupina | C.crupinastrum | Therophite |
| | Echinops | E.cephalotes | Therophite |
| | Helichrysum | H.araxinum | Hemicriptophite |
| | Helichrysum | H.rubicundum | Hemicriptophite |
| | Heteropappus | H.altaicus | Hemicriptophite |
| | Kolepinia | K.teniuisima | Therophite |
| | Lactuca | L.scarioloides | Therophite |
| | Lasiogon | L.muscoides | Therophite |
| | Onopordon | O.leptolepis | Hemicriptophite |
| | Senecio | S.vernalis | Therophite |
| | Sonchus | S.oleraceus | Hemicriptophite |
| | Taraxacum | T.vulgare | Therophite |

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| | Tragopogon | T.buphthalmoides | Therophite |
|---------|---------------|--------------------|-----------------|
| | Tragopogon | T.caricifolium | Therophite |
| | Tragopogon | T.graminifolius | Hemicriptophite |
| | Xanthium | X.spinosum | Hemicriptophite |
| Poaceae | Aegillops | A.tauschii | Therophite |
| | Agropyron | A.longe- aristatum | Hemicriptophite |
| | Alopecurus | A.arandinaceus | Hemicriptophite |
| | Bromus | B.danthonia | Therophite |
| | Bromus | B.tectorum | Therophite |
| | Cynodon | C.dactylis | Therophite |
| | Dactylis | D.glomerata L. | Therophite |
| | Echinochloa | E.crus – galli | Therophite |
| | Eremopyrum | E.confusum | Therophite |
| | Eremopyrum | E.distans | Therophite |
| | Halopyrum | H.muaronatum | Hemicriptophite |
| | Hordeum | H.glaucum | Therophite |
| | Lolium | L.persicum | Therophite |
| | Phleum | Ph.paniculatum | Therophite |
| | Phleum | Ph.pretense | Hemicriptophite |
| | Phragmites | Ph.communis | Hemicriptophite |
| | Poa | P.bulbosa | Hemicriptophite |
| | Setaria | S.viridis | Therophite |
| | Stipa | S.pulcherrima | Hemicriptophite |
| | Taeniatheriun | T.crinitum | Therophite |
| | Trachynia | T.distachya | Therophite |

Conclusion

In this research dominant plants were detected. They were consisting of these families: Asteraceae, Brassicaceae, Fabaceae, Poaceae, Boraginaceae and Polygonaceae. Determining the condition of a plant community is increasingly important as vegetation responds to anthropogenic stress, exotic species invasions, abiotic disturbances, and new management approaches (e.g., Godefroid and Koedam 2003, Abella and Covington 2004). Through their architecture, physiology, growth, and phenology, dominant plants determine overall community structure, such as biomass and canopy strata (Richards 1996) and ecosystem engineering (Malmer et al. 2003); A dominant species can be highly competitive in a core habitat or able to tolerate stress in a peripheral habitat (Wisheu and Keddy 1992). It can make up a majority of stems in a plot or less than the majority. Species richness can also vary with different dominants (Denslow and Hughes 2004). A particular species can vary in its dominance or dominate wherever it occurs (Lavoie et al. 2003). A few authors characterize dominant plants in relation to the number of co-occurring species. Some of scientists call those that coexist with many species. such as alpine tundra sedges, "conservative dominants." (Theodose and Bowman 1997)In contrast, some of them (Hodgson et al. 1998) described abundant plants of speciespoor assemblages as "aggressive dominants." Invasive or transformer species (Richardson et al. 2000), such as Phalaris arundinacea (reed canarygrass) and Typha x glauca (hybrid cattail), behave in this way, tending to exclude other species and create monotypic stands (Galatowitsch et al. 1999). While not quantitative, these distinctions begin to address the different roles and behaviors of dominant species. The presence or abundance of invasive species has also been suggested as an indicator of wetland quality. However, (Denslow and Hughes 2004) note that complex community interactions can allow a blurring of the distinction between native and exotic dominants, as native dominants become management issues and exotic dominants do not always decrease species diversity.

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